

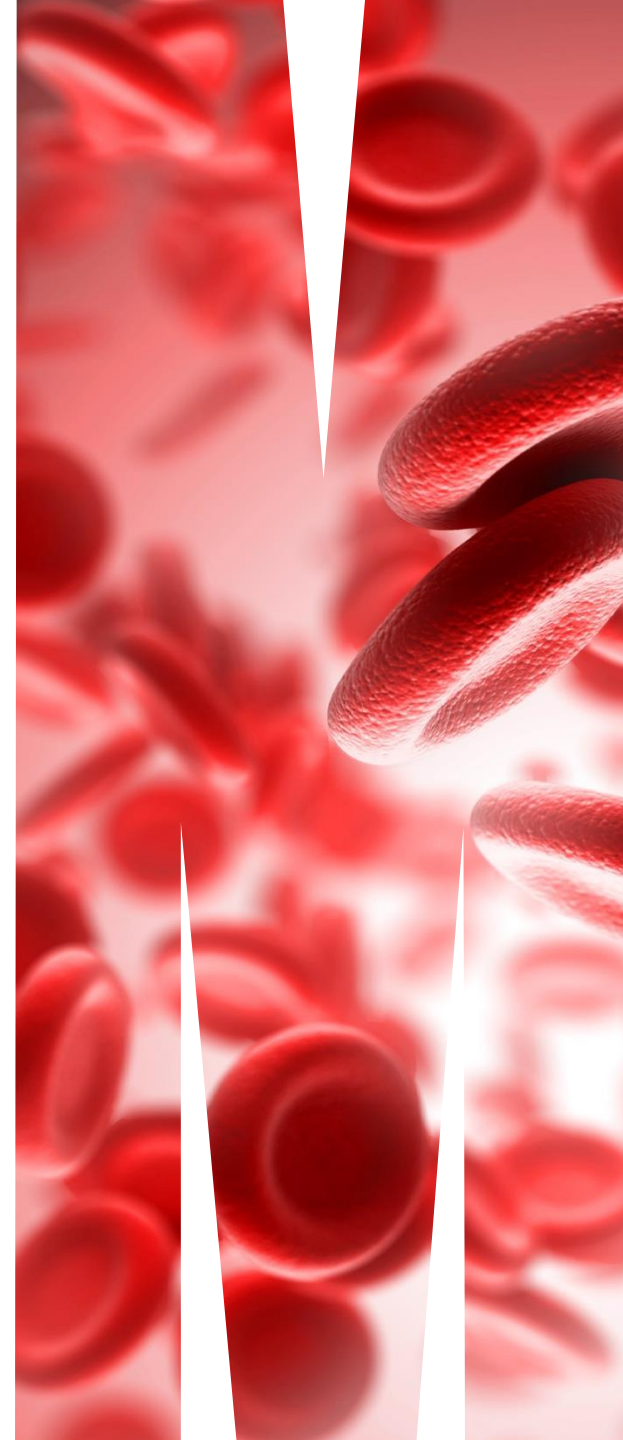
MPN Education Symposium

What is an MPN in 2025

Andrew Perkins

- Professor of Haematology, The Alfred Hospital
- Head of Blood Cancer Genomics, Australian Centre for Blood Diseases, Central Clinical School, Monash University
- Director of Molecular Pathology, Alfred Health

Perth, Oct 25th, 2025



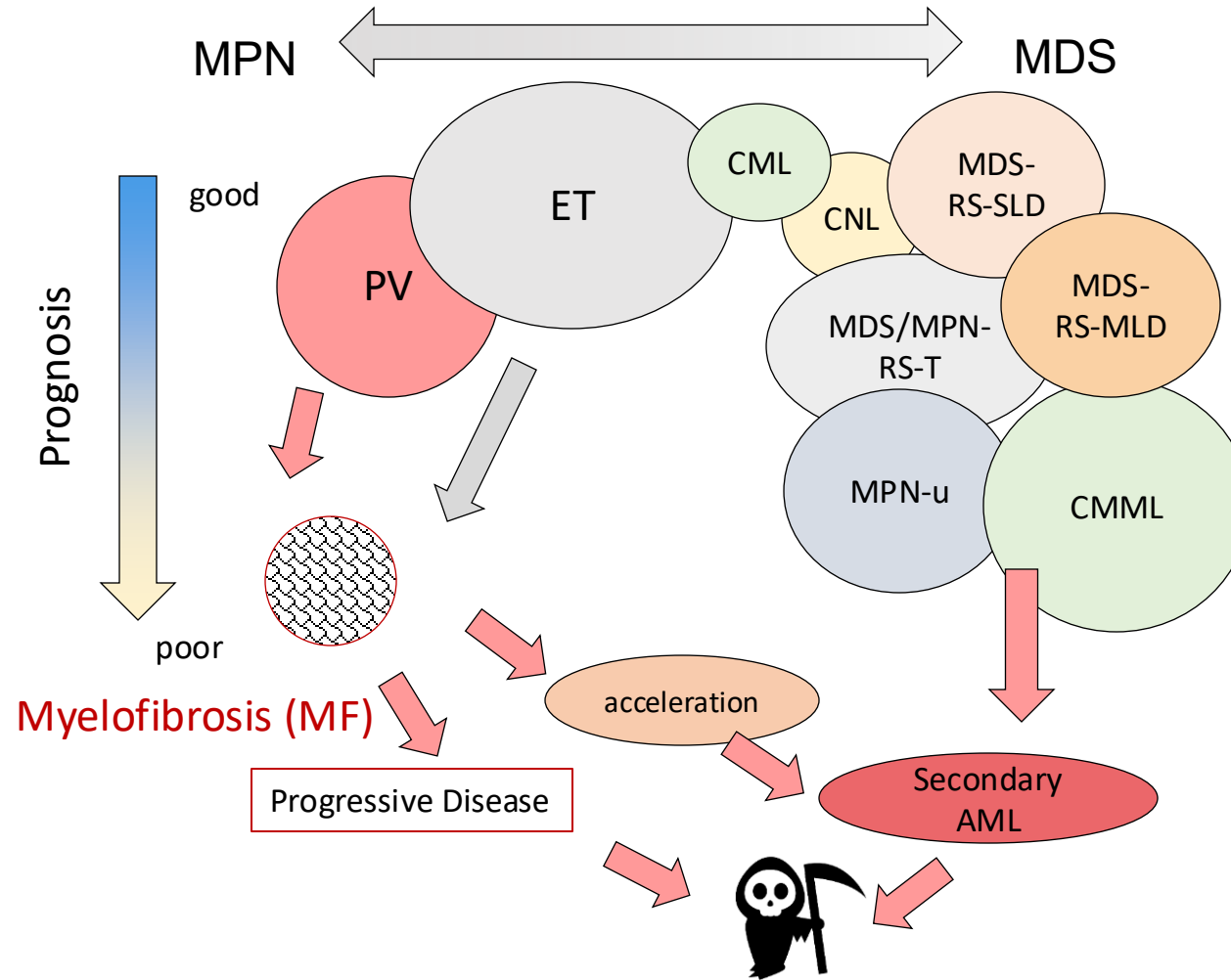
Talk Outline

- What are MPNs
- Haematopoietic Cytokine Signalling 101
- NGS in MPNs: Diagnosis, Prognosis and Discovery
- Germline risk of MPN
- Unmet needs in MPN – a couple of current and future clinical trials

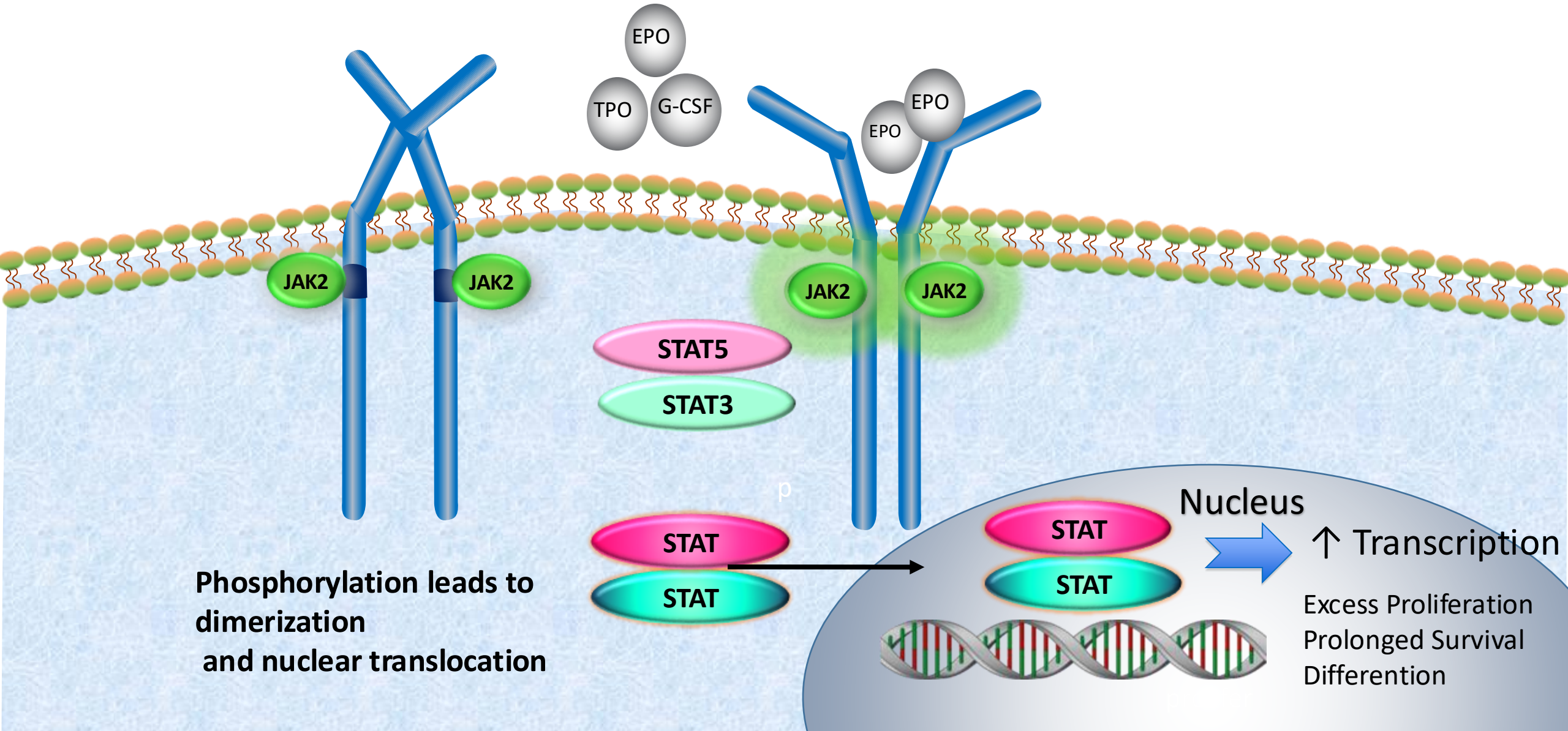
What are MPNs?

Excess Blood
Production

Impaired Blood
Differentiation



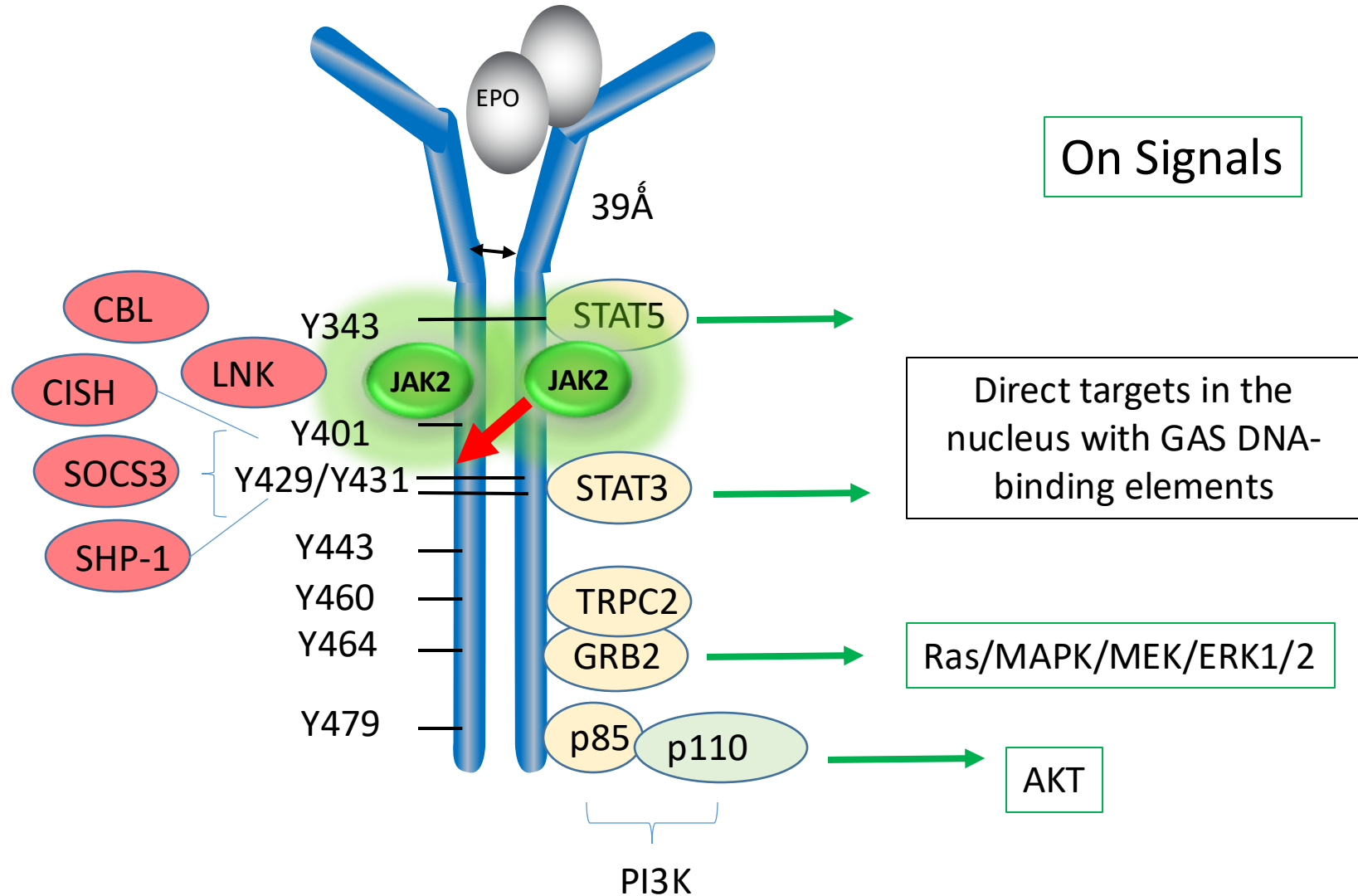
JAK-STAT signalling from type 1 cytokine receptors



Designing a targeted NGS panel from the ground up: Consider EPO and MPL receptor Signalling

Off Signals

On Signals

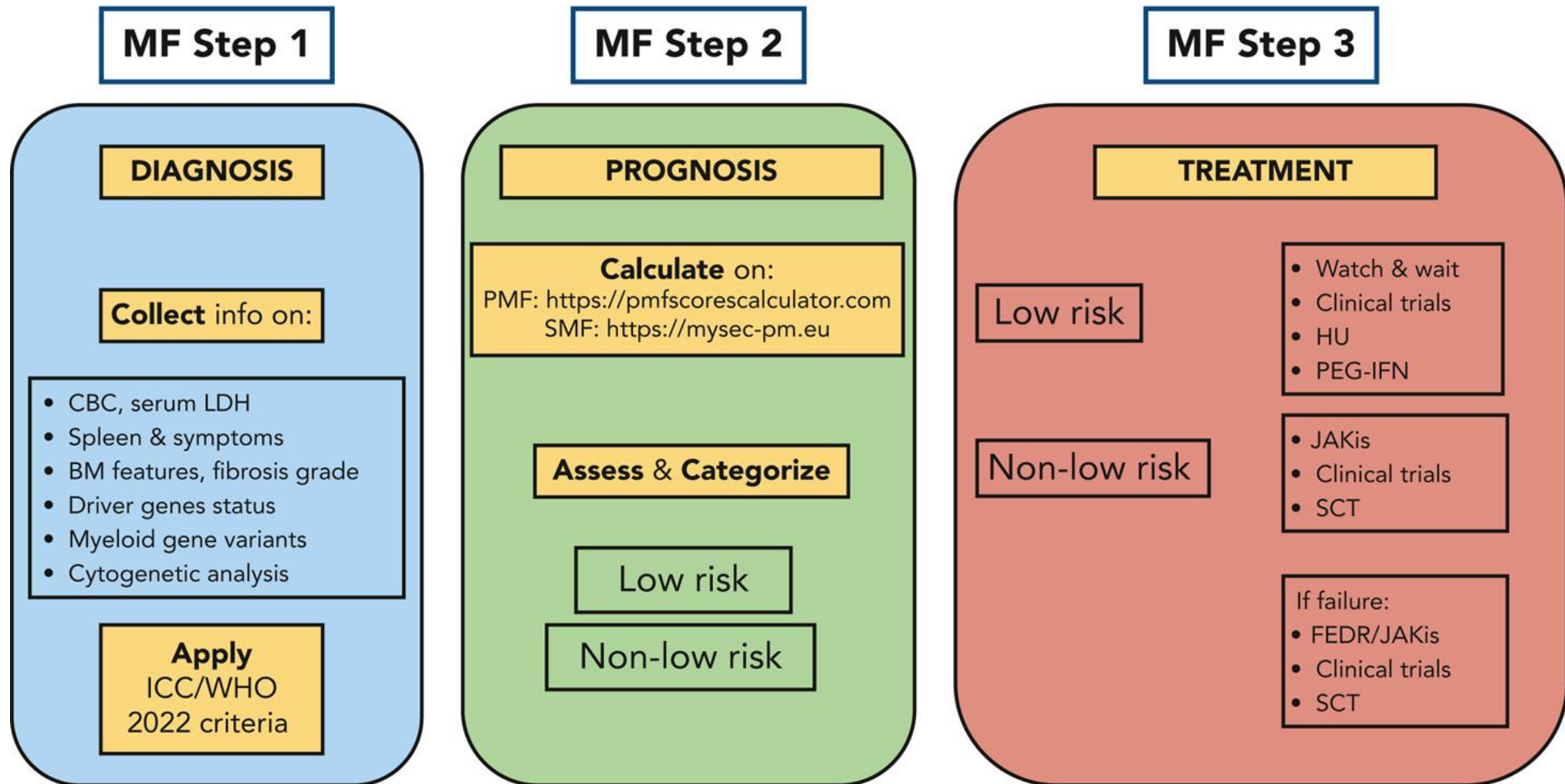


What might we want to use Next Generation Sequencing (NGS) for?

- Diagnosis – increasingly important for WHO classification of all myeloid malignancies
- Prognosis
- Personalised Treatment; e.g. trial options
- Clonal architecture and evolution/progression
- Minimal Residual Disease Monitoring
- Discovery



The 3 steps of MF management



Legend: BM= bone marrow; CBC= complete blood count; FEDR= fedratinib, HU= hydroxyurea; ICC= International Consensus Classification; JAKis= JAK inhibitors; LDH= lactate dehydrogenase; MF= myelofibrosis; PEG-IFN= PEGylated-Interferon; PMF= primary myelofibrosis; SCT= stem cells transplantation; SMF= secondary myelofibrosis; WHO= World Health Organization.

Francesco Passamonti, Barbara Mora, Myelofibrosis, Blood, 2023,

Copyright © 2023 American Society of Hematology

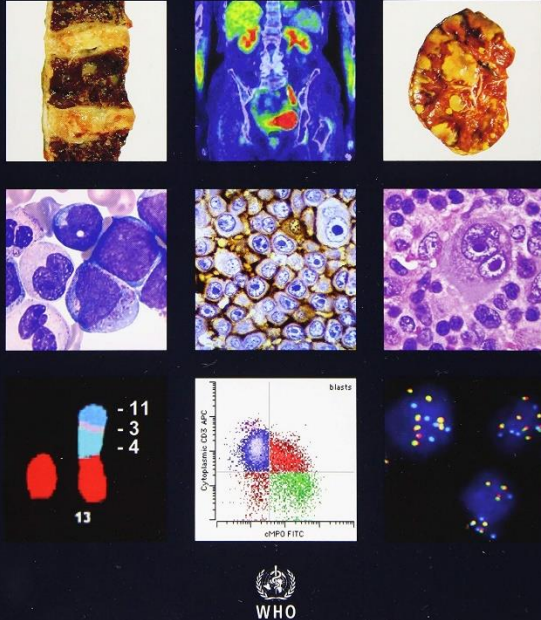


American Society of Hematology
Helping hematologists conquer blood diseases worldwide

Diagnosis of MPNs

WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues

Steven H. Swerdlow, Elias Campo, Nancy Lee Harris, Elaine S. Jaffe, Stefano A. Pileri, Harald Stein, Jürgen Thiele, Daniel A. Arber, Robert P. Hasserjian, Michelle M. Le Beau, Attilio Orazi, Reiner Siebert



Histology

Imaging

Cytogenetics

FISH

FACS

Molecular Testing

Pathologists love to put things in boxes

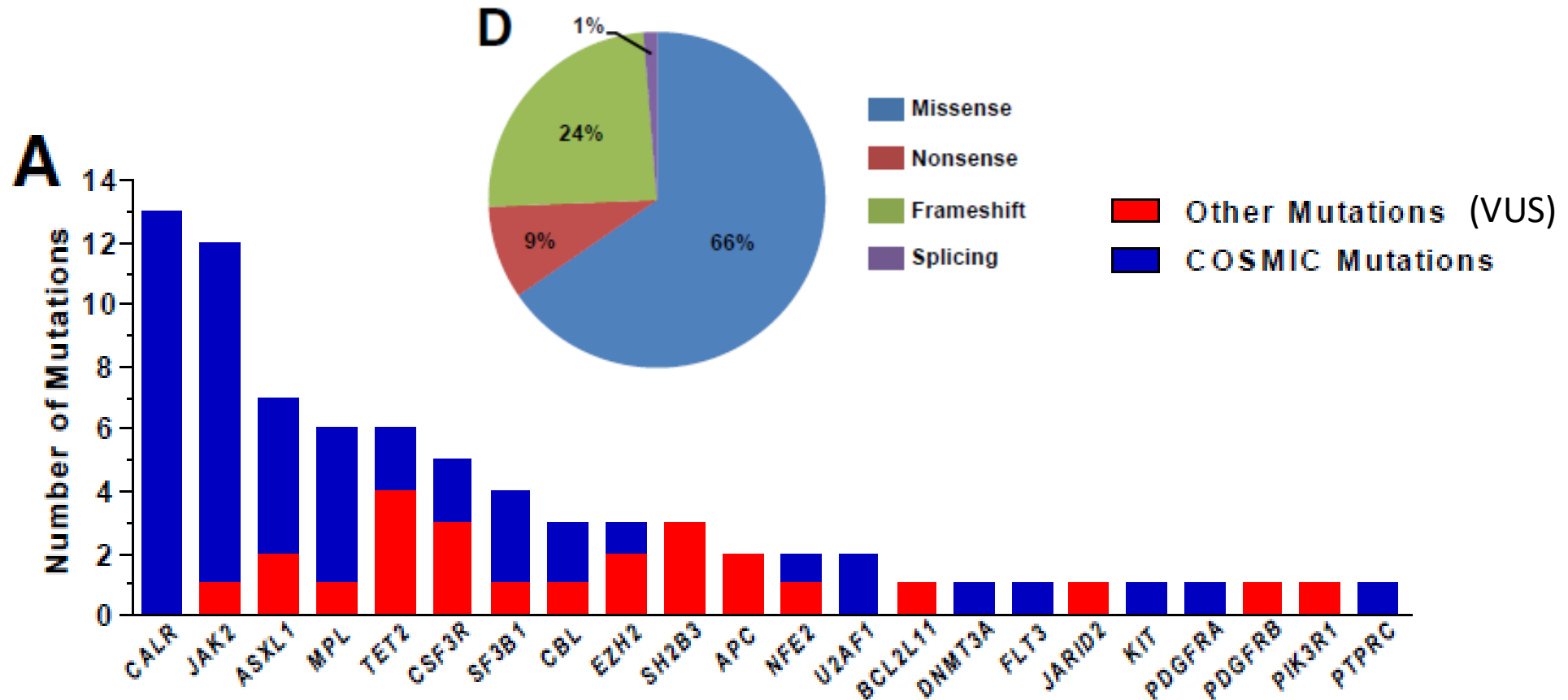




Rapid Molecular Profiling of Myeloproliferative Neoplasms Using Targeted Exon Resequencing of 86 Genes Involved in JAK-STAT Signaling and Epigenetic Regulation



Graham W. Magor,^{*} Michael R. Tallack,^{*} Nathan M. Klose,[†] Debra Taylor,[‡] Darren Korbie,^{§¶} Peter Mollee,^{||} Matt Trau,^{§¶**} and Andrew C. Perkins^{*||††}



What Genes are important in MPNs?

- 2035 MPNs (ET, PV, MF [309], other [49])
- 69 genes including CNV
- >200 WES

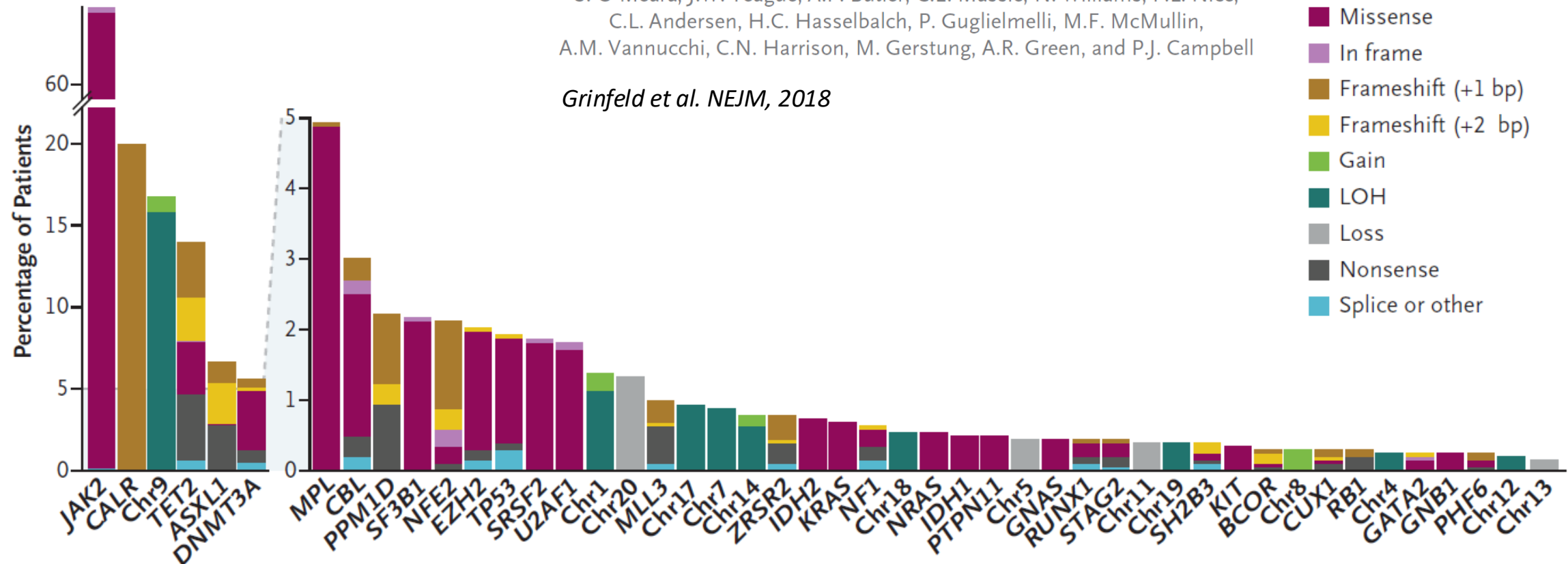
ORIGINAL ARTICLE

Classification and Personalized Prognosis in Myeloproliferative Neoplasms

J. Grinfeld, J. Nangalia, E.J. Baxter, D.C. Wedge, N. Angelopoulos, R. Cantrill, A.L. Godfrey, E. Papaemmanuil, G. Gundem, C. MacLean, J. Cook, L. O'Neil, S. O'Meara, J.W. Teague, A.P. Butler, C.E. Massie, N. Williams, F.L. Nice, C.L. Andersen, H.C. Hasselbalch, P. Guglielmelli, M.F. McMullin, A.M. Vannucchi, C.N. Harrison, M. Gerstung, A.R. Green, and P.J. Campbell

Grinfeld et al. NEJM, 2018

A Recurrently Mutated Genes and Chromosomes



Australian Recommendations for NGS in MPNs

Pathology (April 2021) 53(3), pp. 339–348

MYELOID NEOPLASMS

Myeloid somatic mutation panel testing in myeloproliferative neoplasms

DAVID M. ROSS^{1,2,3,4}, CANDICE THOMSON^{1,2}, NADA HAMAD^{1,5},
STEVEN W. LANE^{1,6,7}, KATE MANOS^{1,8}, ANDREW P. GRIGG^{1,8}, BELINDA GUO^{1,9},
WENDY N. ERBER^{1,9,10}, ASHLEIGH SCOTT^{1,6}, NICK VIALLA^{1,11}, LYNETTE CHEE^{1,12},
MAYA LATIMER^{1,13}, COURTNEY TATE^{1,14}, CAROLYN GROVE^{1,9,10,15},
ANDREW C. PERKINS^{1,16}, PIERS BLOMBERY^{1,12}

¹*Myeloproliferative Neoplasms Working Party, Australasian Leukaemia and Lymphoma*



Just 17 genes that cover the majority of diagnostic and prognostic requirements

Categories of mutations in chronic myeloid malignancies

Pathways

Some Genes

GoF mutations



LoF mutations



Cytokine Receptor and Signaling

JAK2, CALR, MPL, CSF3R, KIT, EPOR, JAK3, NRAS, KRAS



Negative regulators of cytokine signalling

SH2B3, CBL, PTPN11



Epigenetics - DNA methylation

TET2, DNMT3A, IDH1, IDH2



Epigenetics - histone modification

EZH2, BCOR, IDH1, IDH2



Splicing

SF3B1, U2AF1, SRSF2, ZRSR2



Transcription

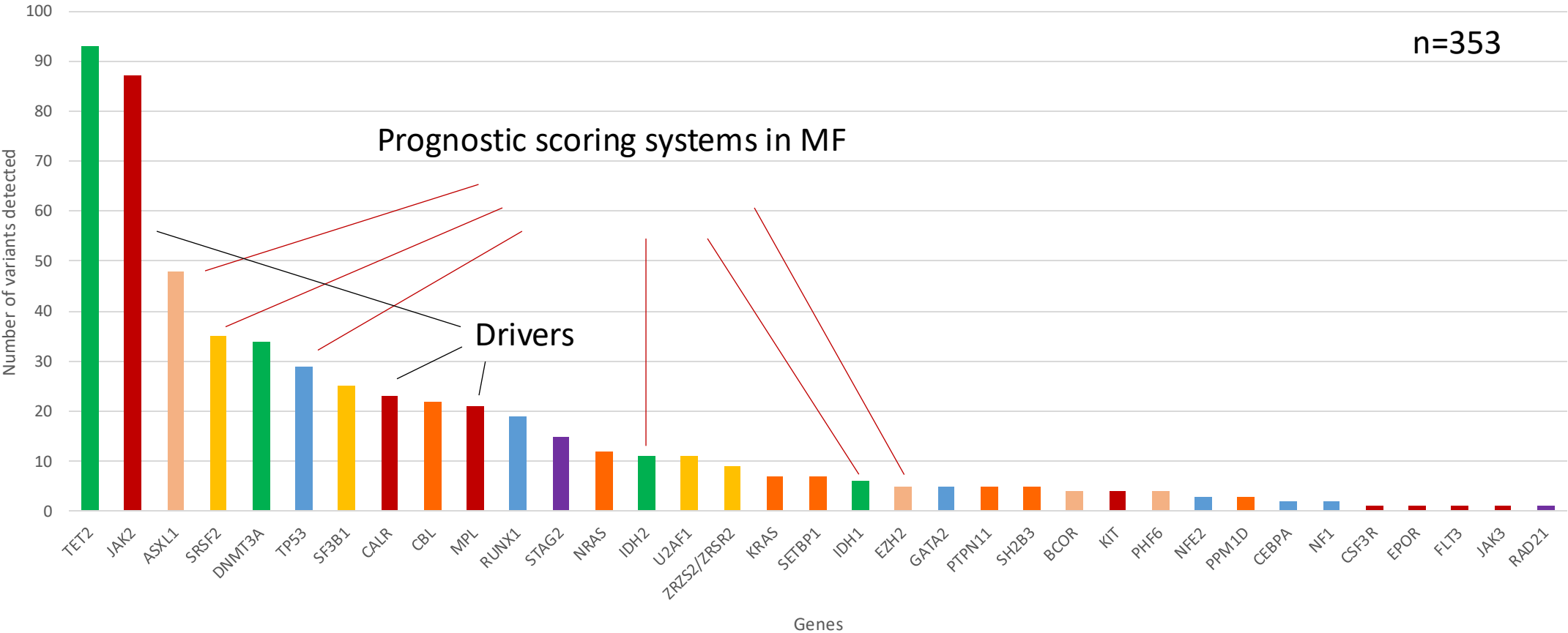
TP53, RUNX1, GATA2, NFE2, CEBPA



Cohesin Complex

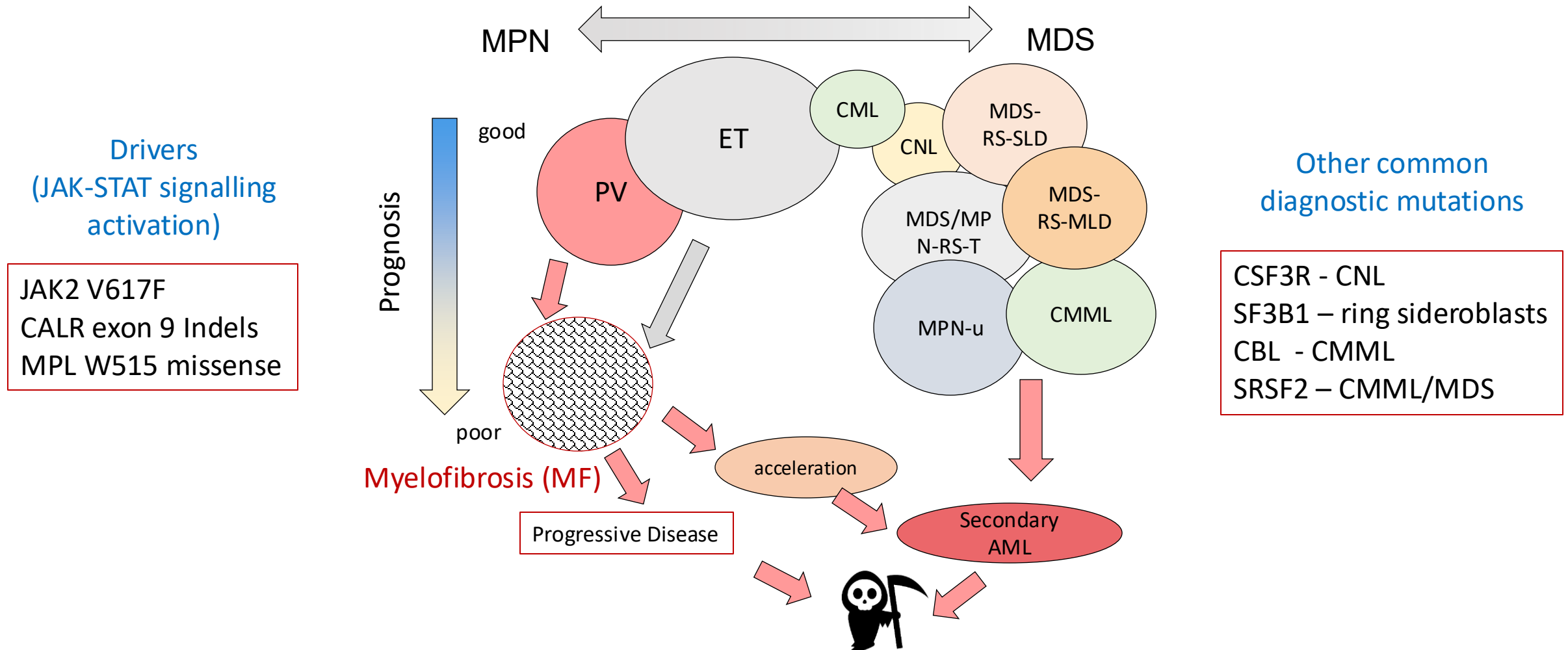
STAG1, STAG2, RAD21

NGS results in chronic myeloid malignancy (2020-2021): Prognosis

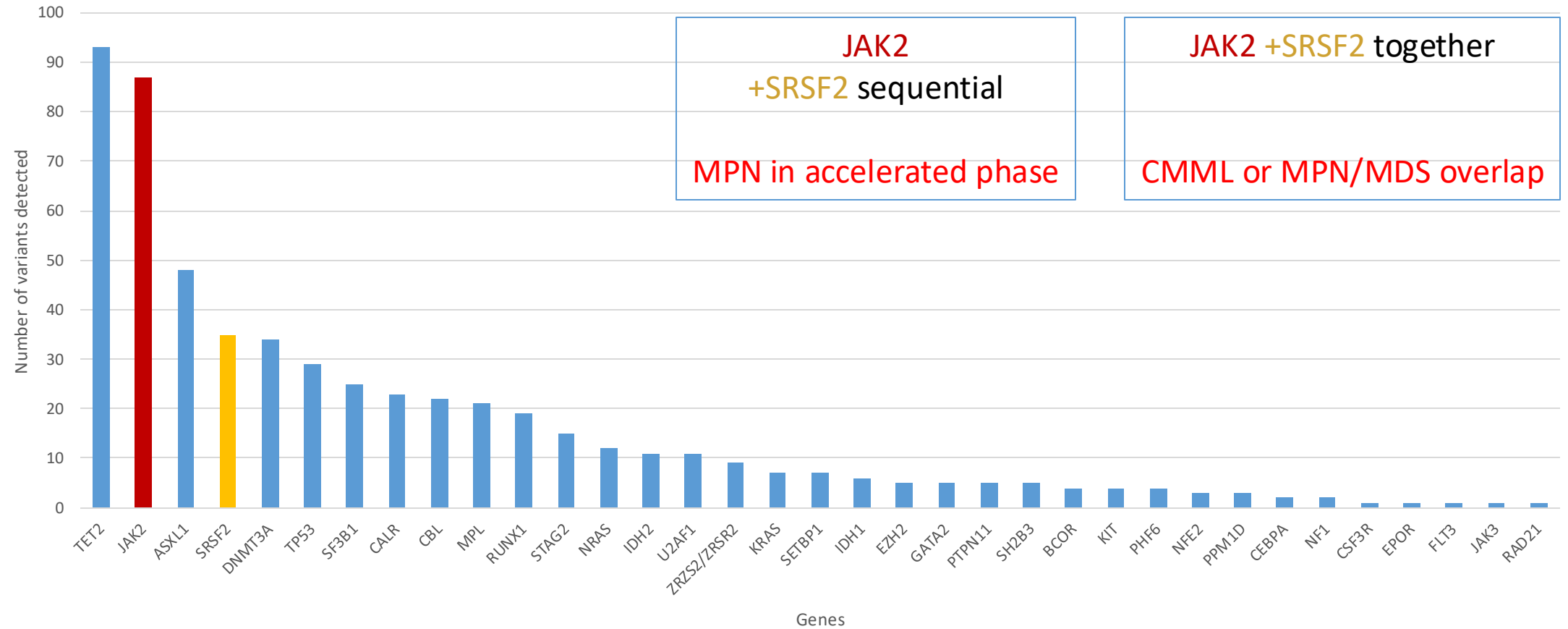


Justin Ng

NGS for Diagnosis

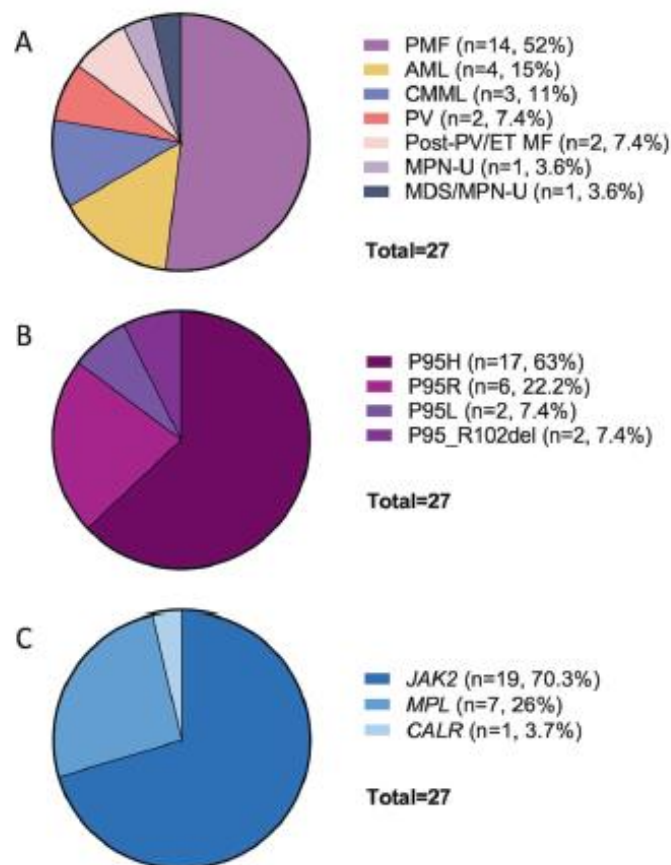


Combinations of mutations result in specific disease entities

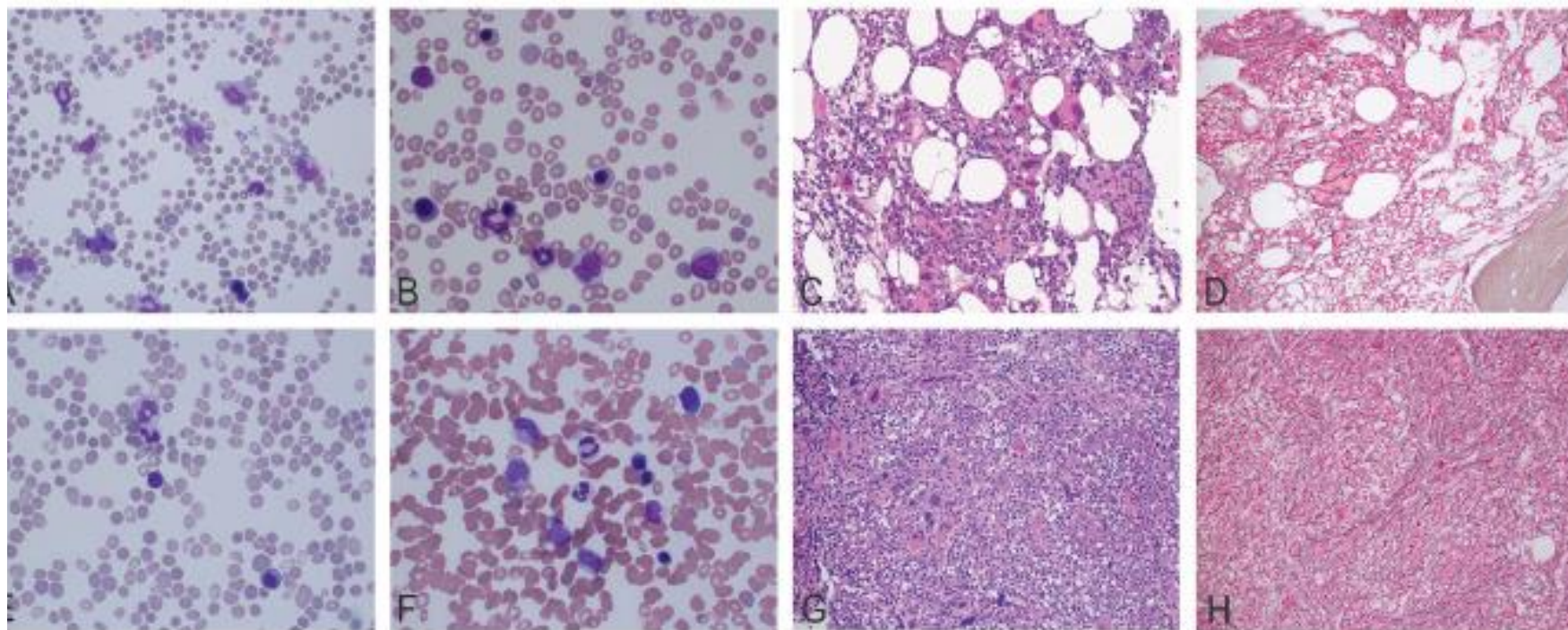


Clinicopathologic spectrum of myeloid neoplasms with concurrent myeloproliferative neoplasm driver mutations and *SRSF2* mutations

Mehrnoosh Tashakori¹, Joseph D. Khoury¹, Mark J. Routbort¹, Keyur P. Patel¹, Sa A. Wang¹, Chi Young OK¹, Siba El-Hussein¹, Rashmi Kanagal-Shamanna¹, Rajyalakshmi Luthra¹, Shimin Hu¹, Pei Lin¹, Naveen Pemmaraju², Prithviraj Bose², Srdan Verstovsek¹, Carlos E. Bueso-Ramos¹, L. Jeffrey Medeiros¹ and Sanam Loghavi¹✉



PMF with monocytosis



CMMML with MF3

PROGNOSIS

MIPSS70 score Home Credits

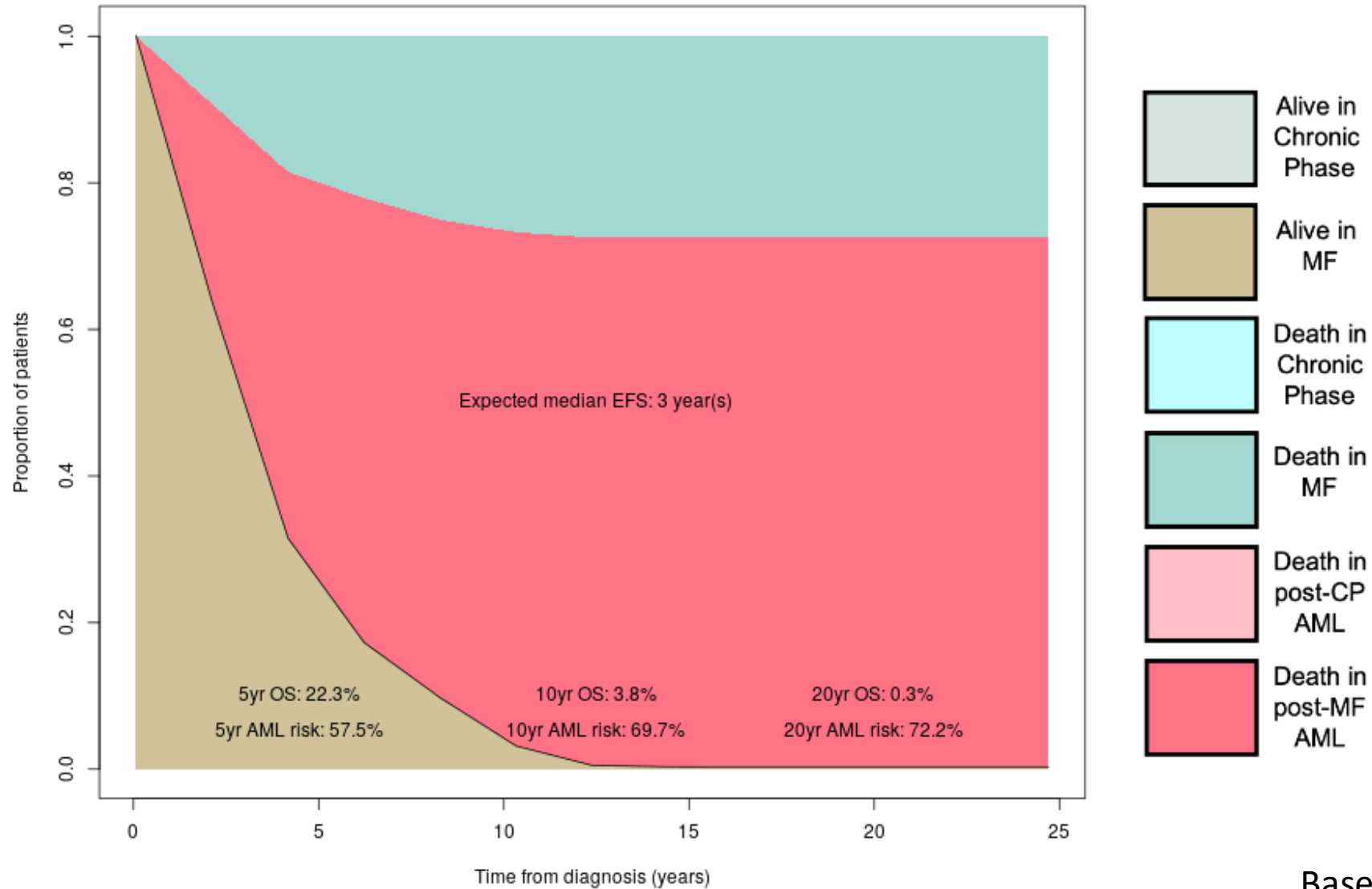
#	Question	Answer
1	Severe Anemia (hemoglobin <80g/L)	☐ Yes ☐ No
2	Moderate Anemia (hemoglobin 80-100g/L)	☐ Yes ☐ No
3	Leucocytosis >25x10 ⁹ /L	☐ Yes ☐ No
4	Thrombocytopenia (platelet count <100x10 ⁹ /L)	☐ Yes ☐ No
5	Peripheral blood blast count ≥2%	☐ Yes ☐ No
6	Bone marrow fibrosis grade ≥2	☐ Yes ☐ No
7	Constitutional symptoms	☐ Yes ☐ No
8	Absence of CALR type 1/like mutation	☐ Yes ☐ No
9	HMR ¹ category	☐ Yes ☐ No
10	≥2 HMR mutated genes	☐ Yes ☐ No
11	Unfavorable karyotype ²	☐ Yes ☐ No ☐ Not available
12	Very High Risk karyotype ³	☐ Yes ☐ No ☐ Not available

} IHD1, IDH2, SRSF2, ASXL1, U2AF1

Score	Result
MIPSS70	

MPN report patient survival diagram

<https://www.sanger.ac.uk/science/tools/progmod/progmod/>



Based on Grinfeld et al. NEJM 2018

Article

Inherited myeloproliferative neoplasm risk affects haematopoietic stem cells

<https://doi.org/10.1038/s41586-020-2786-7>

Received: 2 October 2019

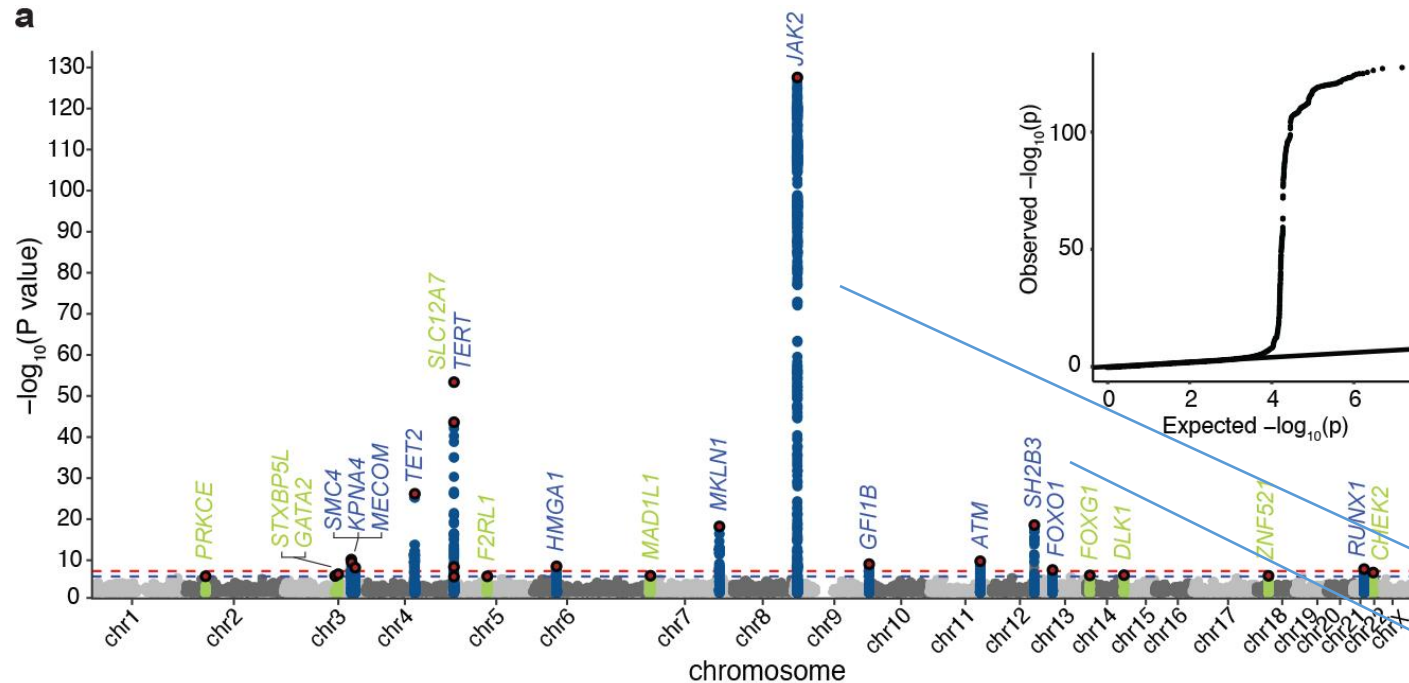
Accepted: 3 July 2020

Published online: 14 October 2020

 Check for updates

Erik L. Bao^{1,2,3,4,59}, Satish K. Nandakumar^{1,2,3,59}, Xiaotian Liao^{1,2,3,59}, Alexander G. Bick^{3,5,6,7,8}, Juha Karjalainen⁹, Marcin Tabaka³, Olga I. Gan^{10,11}, Aki S. Havulinna⁹, Tuomo T. J. Kiiskinen⁹, Caleb A. Lareau^{1,2,3,12}, Aitzkoa L. de Lapuente Portilla¹³, Bo Li^{3,14}, Connor Emdin^{3,5}, Veryan Codd^{15,16}, Christopher P. Nelson^{15,16}, Christopher J. Walker¹⁷, Claire Churchhouse³, Albert de la Chapelle¹⁷, Daryl E. Klein¹⁸, Björn Nilsson^{3,13}, Peter W. F. Wilson^{19,20}, Kelly Cho^{21,22}, Saiju Pyarajan²¹, J. Michael Gaziano^{21,22}, Nilesh J. Samani^{15,16}, FinnGen^{*}, 23andMe Research Team^{*}, Aviv Regev^{3,23,24}, Aarno Palotie^{3,9}, Benjamin M. Neale³, John E. Dick^{10,11}, Pradeep Natarajan^{3,5,25}, Christopher J. O'Donnell^{7,22}, Mark J. Daly^{3,9}, Michael Milyavsky²⁶, Sekar Kathiresan^{3,5,27} & Vijay G. Sankaran^{1,2,3,28,62}

Inherited MPN Risk



a. GWAS meta-analysis for MPNs (n = 2,949 cases and 835,554 controls). X axis is chromosomal position, and y axis is the $-\log_{10}(P)$ value of association (two-tailed, logistic regression). Association signals reaching genome-wide significance ($P < 5 \times 10^{-8}$) and suggestive significance ($P < 1 \times 10^{-6}$) are shown in blue and green, respectively. Red points represent conditionally independent lead variants within each locus.

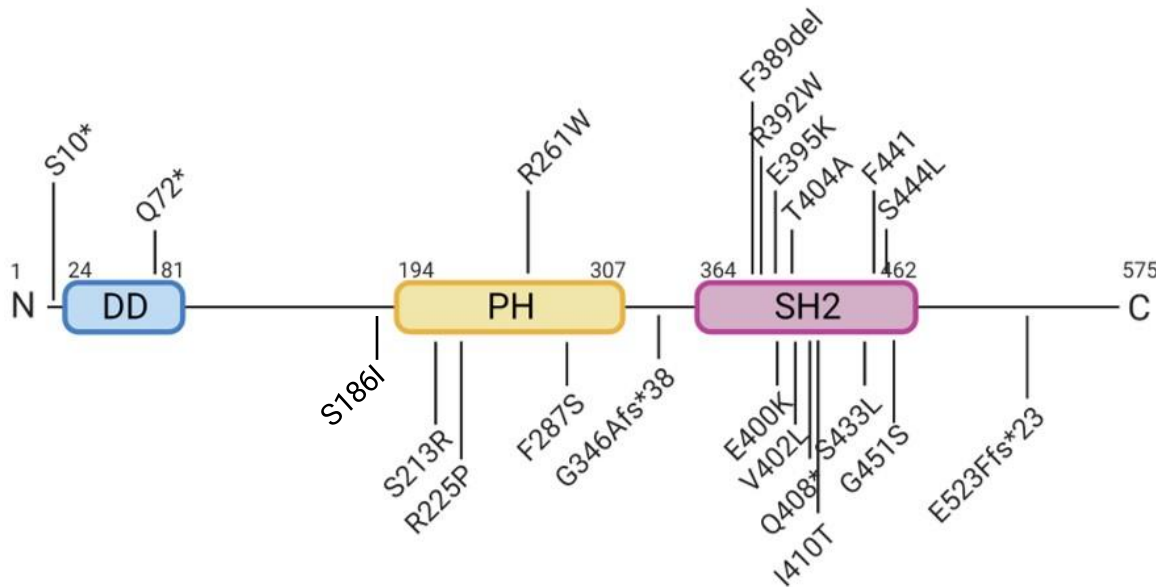
JAK2 46/1 haplotype of four non-coding SNPs

SH2B3

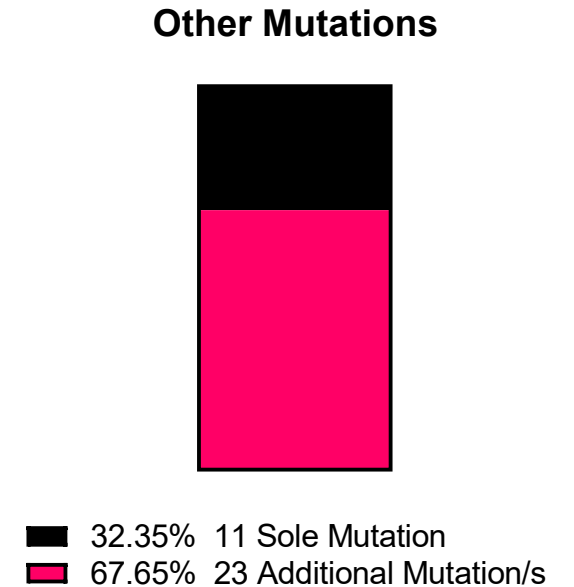
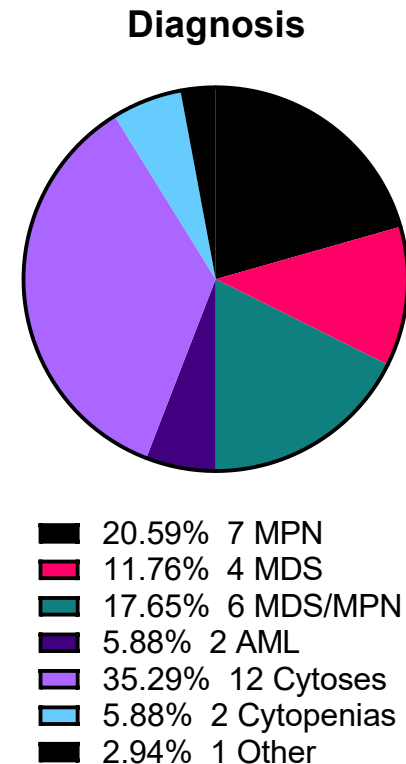
***SH2B3* In Myeloproliferative Neoplasms**

Andrew Perkins Liesl Butler

***SH2B3* variants are frequently missense and in the SH2 domain. Clinical presentations are diverse.**



~2.5% of all Alfred Pathology myeloid next generation sequencing samples are found to have an *SH2B3* variant

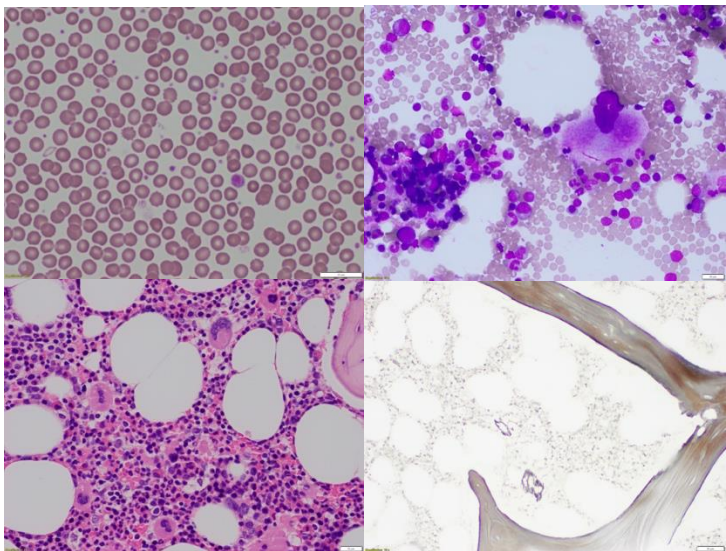


Germline missense *SH2B3* variants may lead to myeloproliferative disease

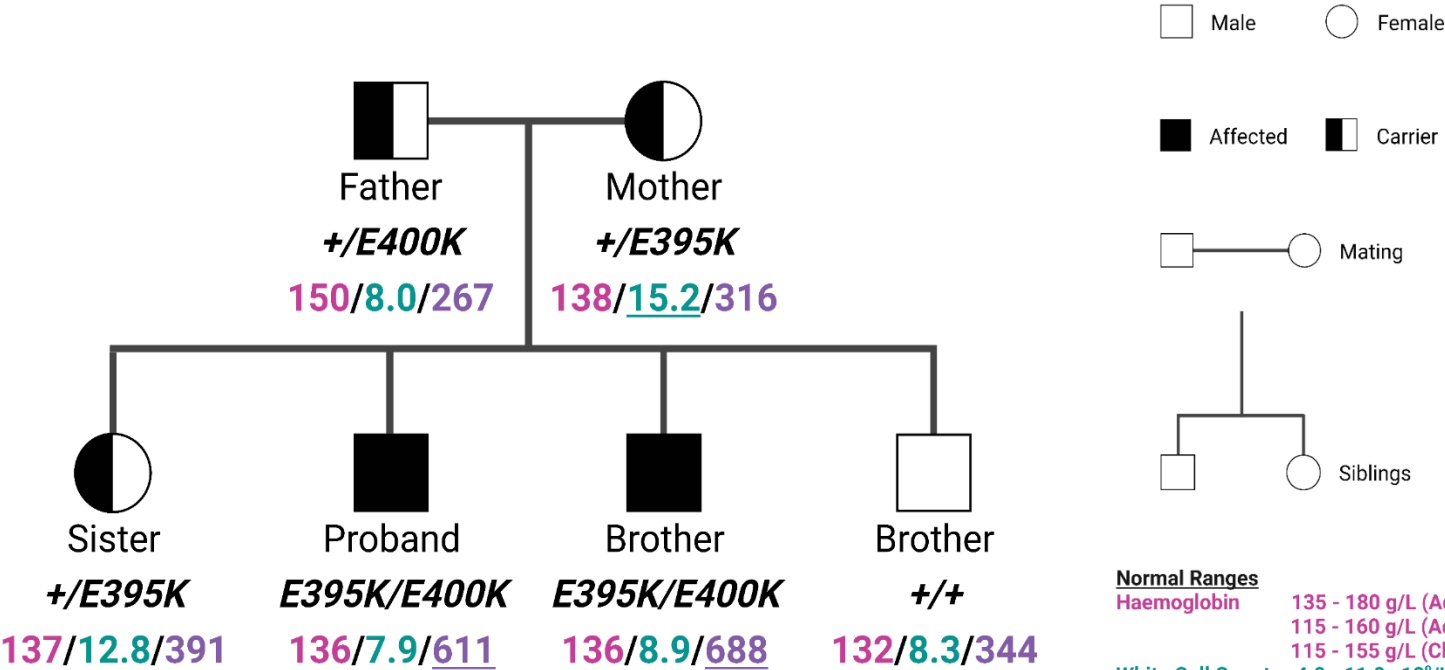
Family 1: E395K

Family 2: E395K/E400K

Family 3: T404A

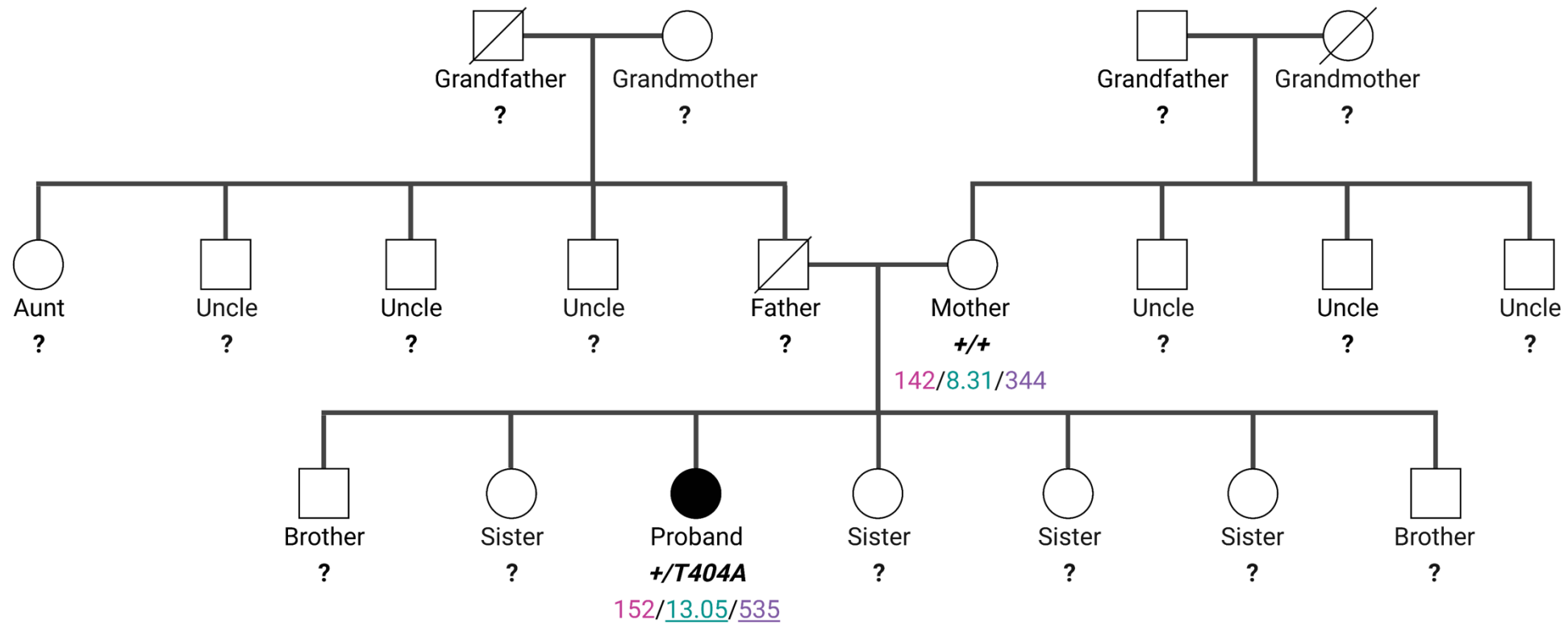


Proband's blood and bone marrow (aspirate, trephine and reticulin stain)

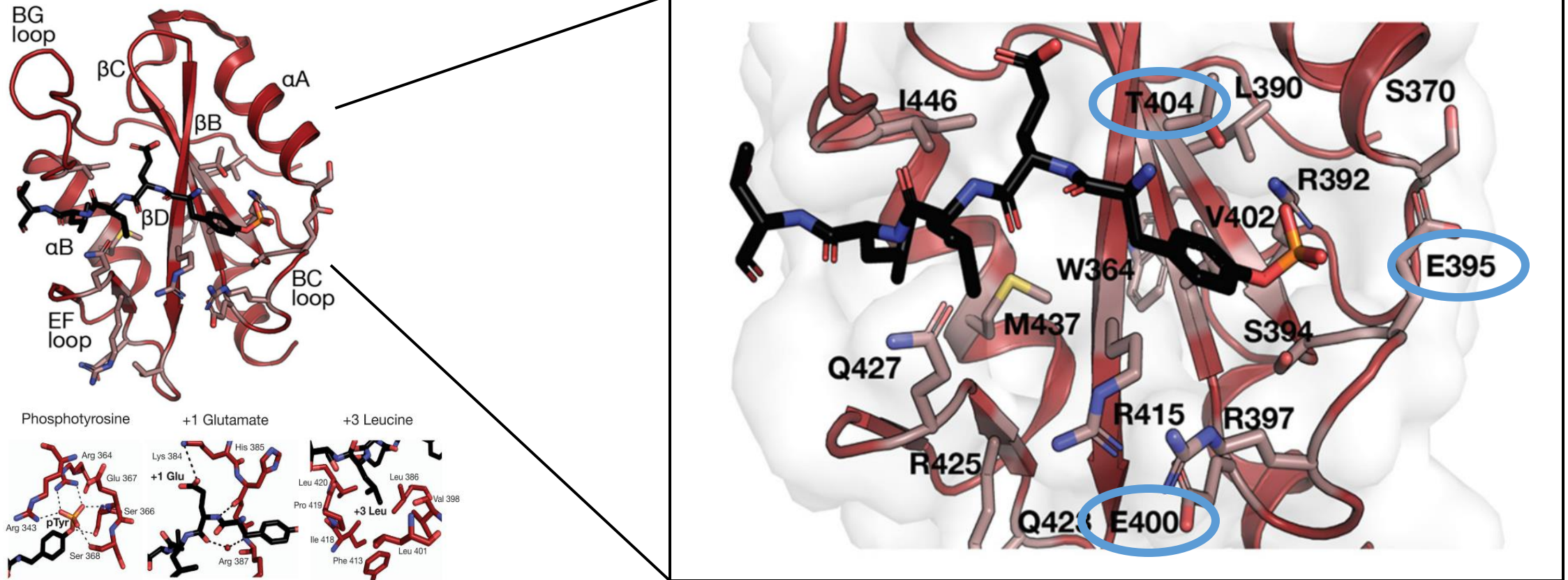


SH2B3-T404A mutations may represent a germline cause for myeloproliferative disease

Family 2: *T404A*



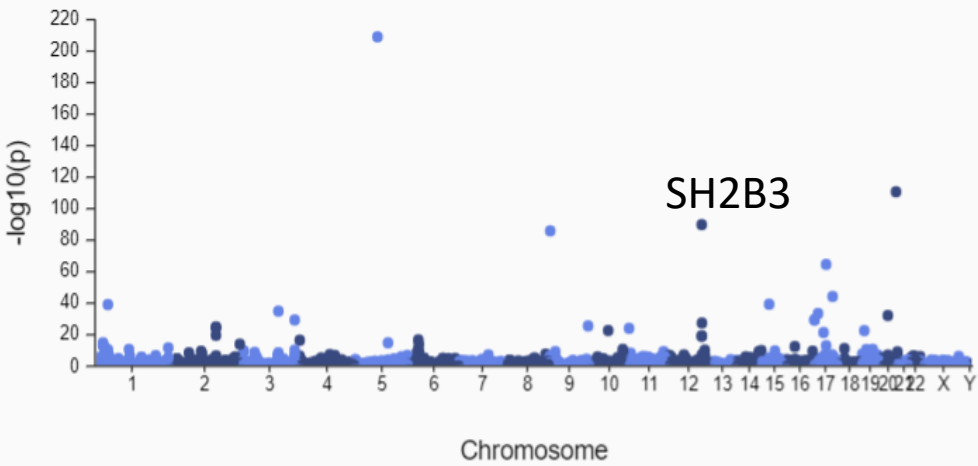
Variants in the SH2 domain of SH2B3 affect interaction with phosphorylated JAK2



(Morris et al. 2021)
(Morris et al. 2022)

Gene: SH2B3 (ENSG00000111252) Phenotype: Platelet count Burden set: ● missense|LC

missense|LC gene burden associations with Platelet count



Burden set

pLoF

missense|LC

synonymous

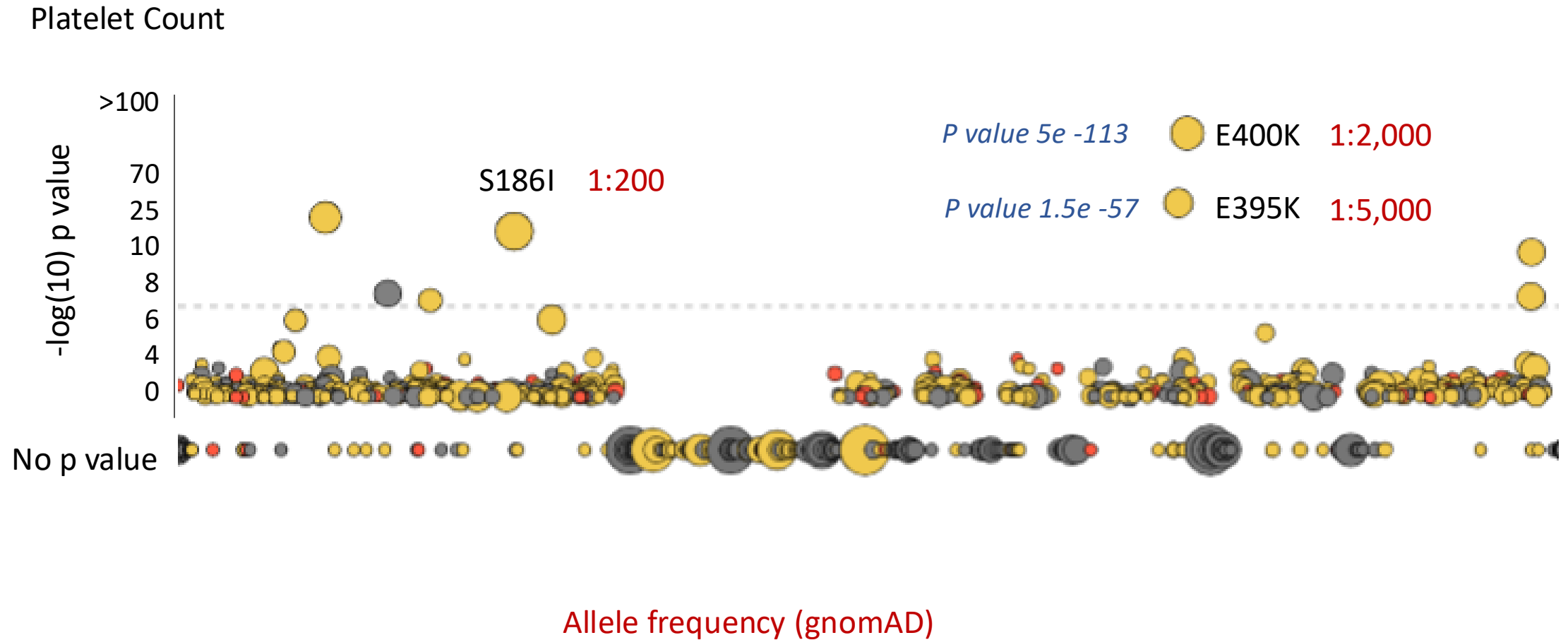
Filter genes

Min P-value

-Infinity

Gene Name	P-Value SKATO	P-Value Burden	P-Value SKAT	BETA Burden	Details
IQGAP2	8.41e-210	3.36e-74	1.24e-198	-5.13e-3	+
TUBB1	1.94e-111	4.6e-53	2.81e-111	-4.76e-3	+
SH2B3	1.36e-90	7.74e-36	2.66e-88	5.14e-3	+
JAK2	1.24e-86	2.58e-65	6.12e-70	7.02e-3	+
ITGA2B	2.36e-65	6.6e-60	1.78e-46	-6.85e-3	+
APOH	5.42e-45	9e-6	7.74e-46	1.38e-3	+

Inherited missense variants in SH2B3 cause thrombocytosis and leukocytosis



Functional Validation: Does *SH2B3-E395K* result in a myeloproliferative disease phenotype?

1
GCA
CAG
CTG
GTC
CAG
CTC
CAG
GGC
CCT
GAT
GCC
CAC
GGC
GTG
TTC
1▶
A
Q
L
V
Q
L
Q
G
P
D
A
H
G
V
F

XbaI

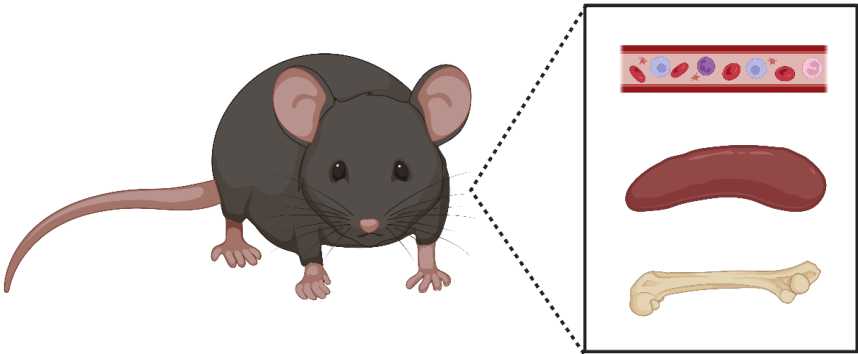
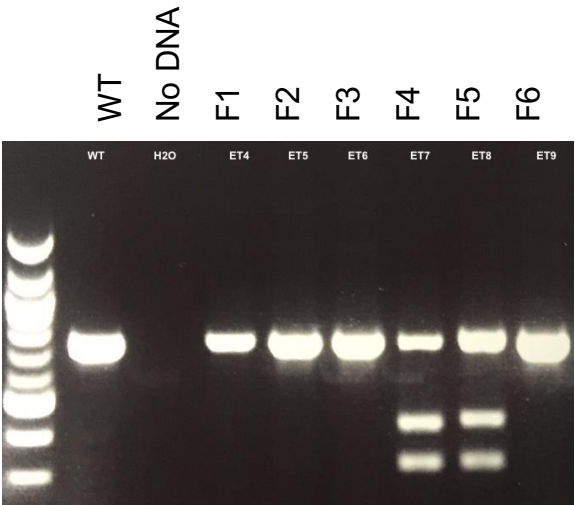
E367K

46
CTG
GTG
CGG
CAG
AGT
AAG
TCT
AGA
AGA
GGA
GAG
TAT
GTA
CTC
ACA
16▶
L
V
R
Q
S
K
S
R
R
G
E
Y
V
L
T

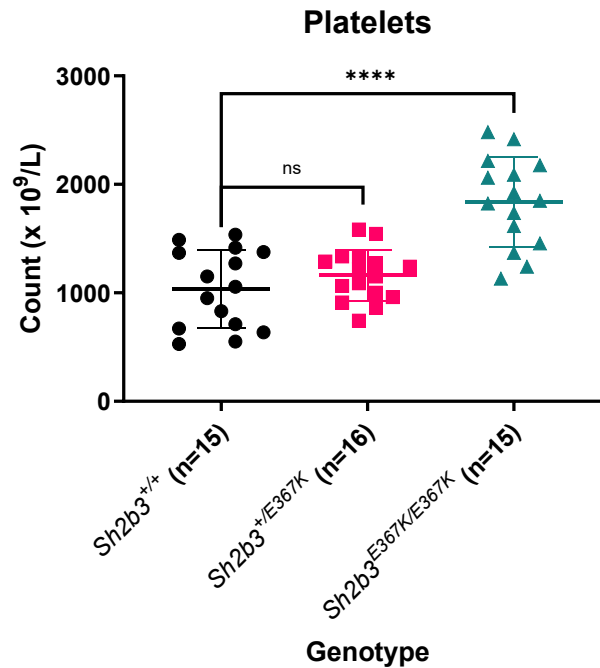
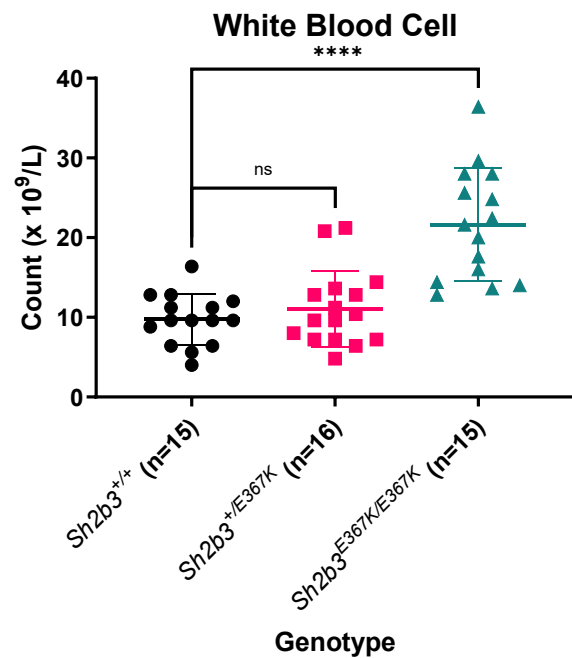
91
TTC
AAC
TTA
CAG
GGC
AGA
GCC
AAG
GTAGGGGCAG
GAACT
31▶
F
N
L
Q
G
R
A
K



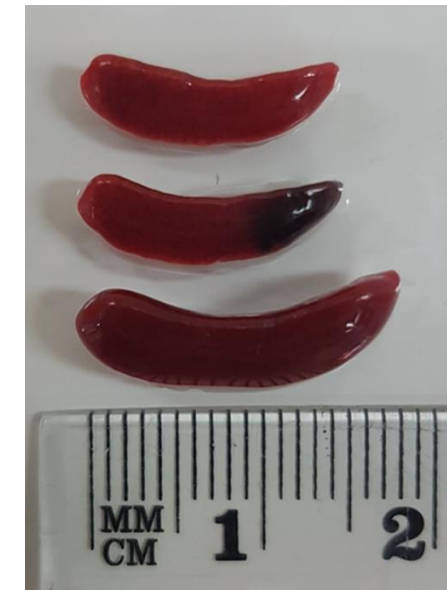
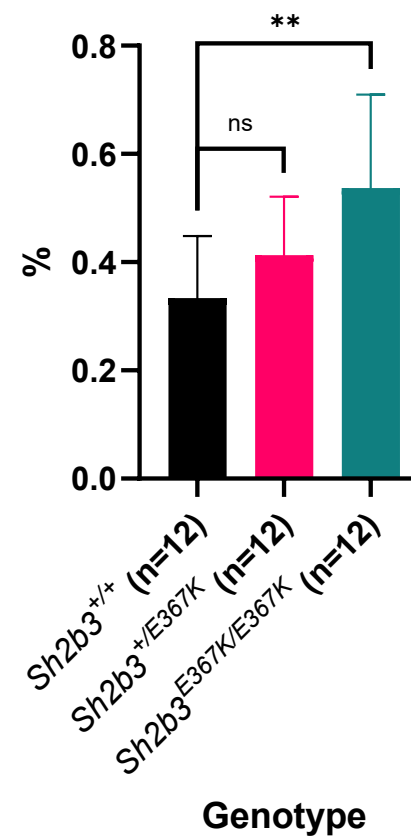
Monash Genome Editing Platform



Sh2b3-E367K mutant mice demonstrate peripheral blood cytoses and splenomegaly

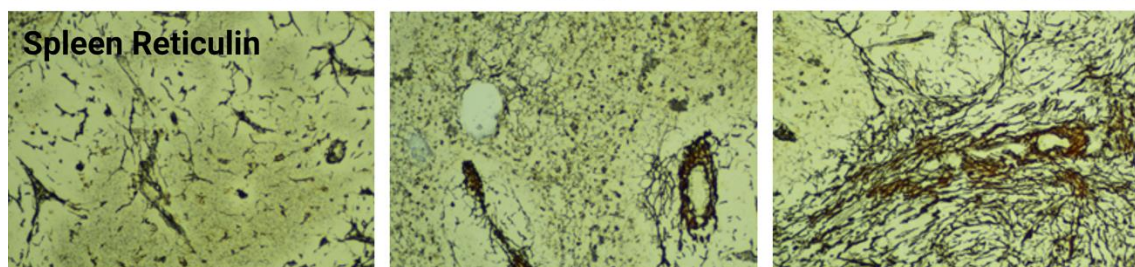
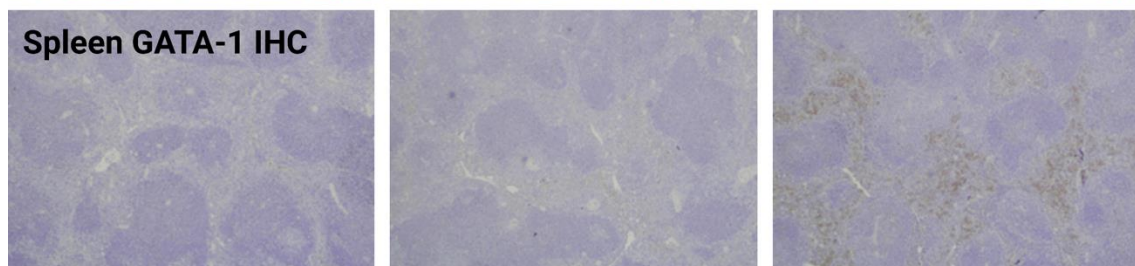
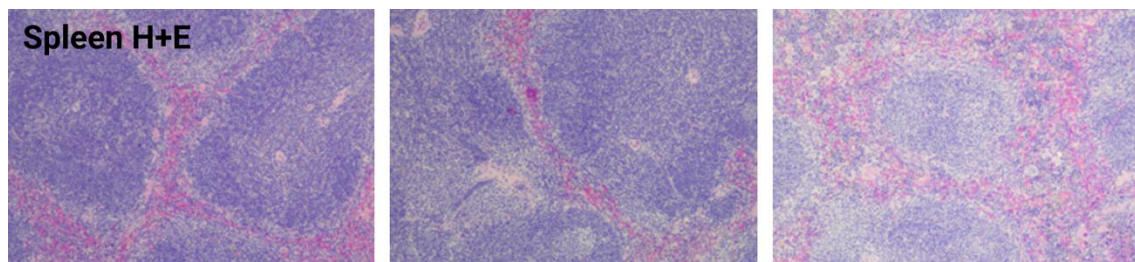


Spleen Of Body Weight



Statistical significance determined using the Mann Whitney test

Sh2b3-E367K mutated mice exhibit reticulin fibrosis

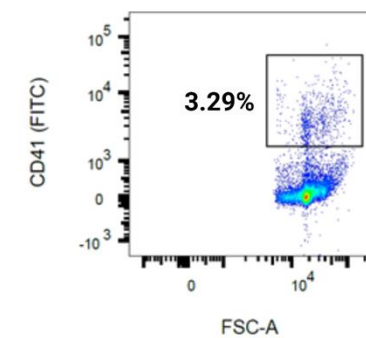
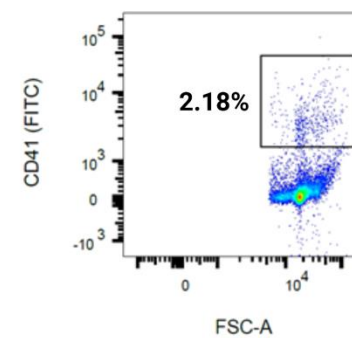
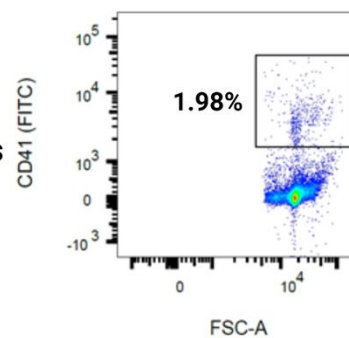
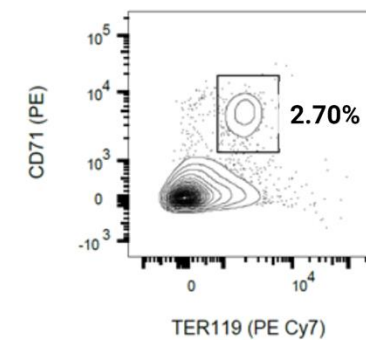
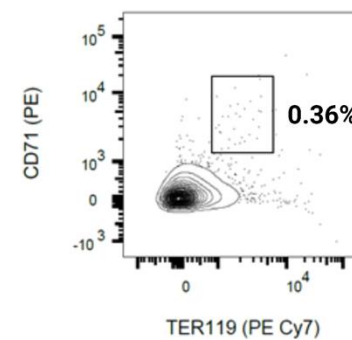
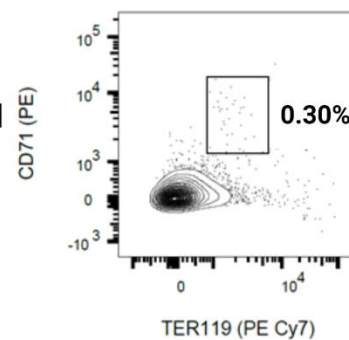


Sh2b3^{+/+}

Sh2b3^{+/E367K}

Sh2b3^{E367K/E367K}

Spleen Erythroid Cells

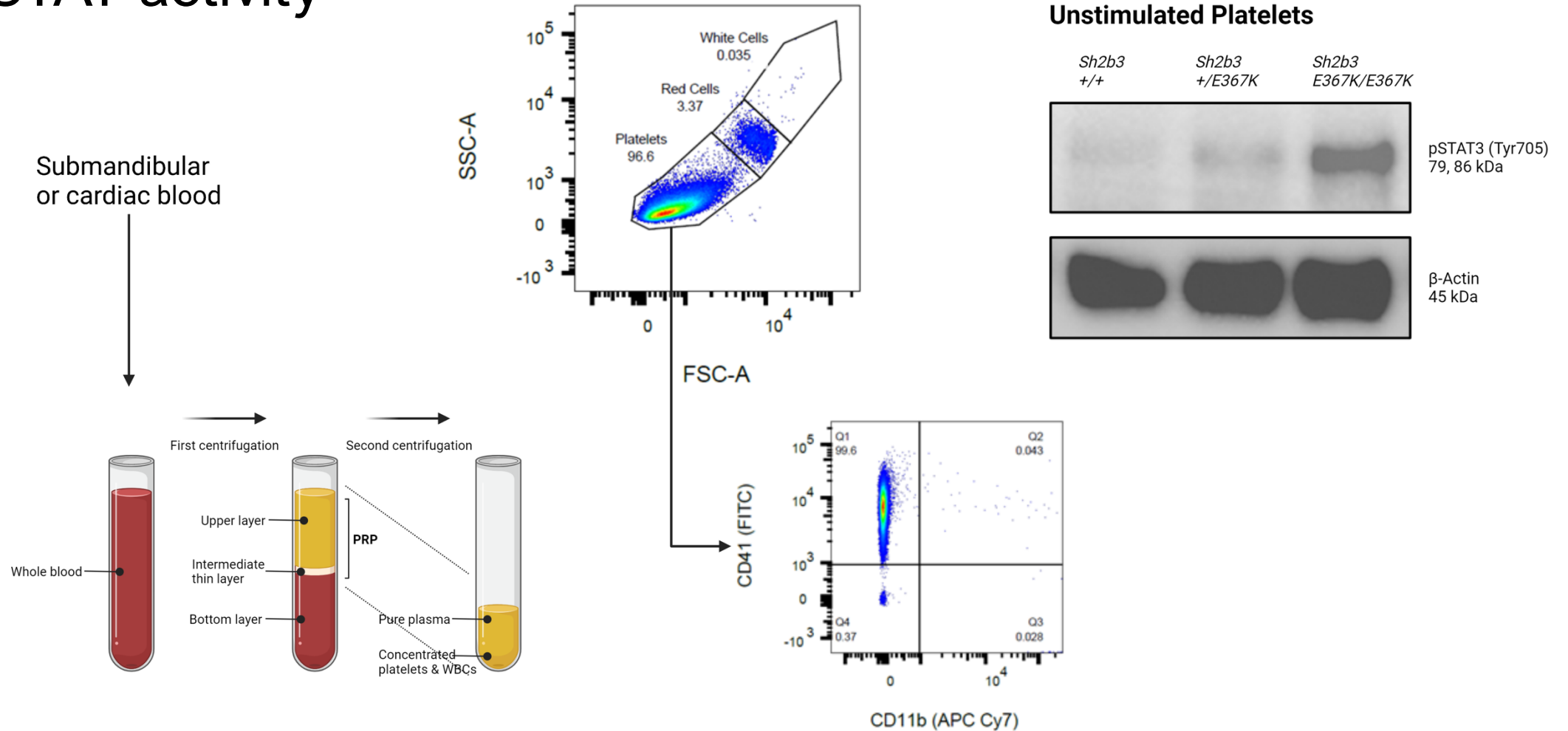


Sh2b3^{+/+}

Sh2b3^{+/E367K}

Sh2b3^{E367K/E367K}

Sh2b3-E367K homozygous mouse platelets have heightened JAK-STAT activity



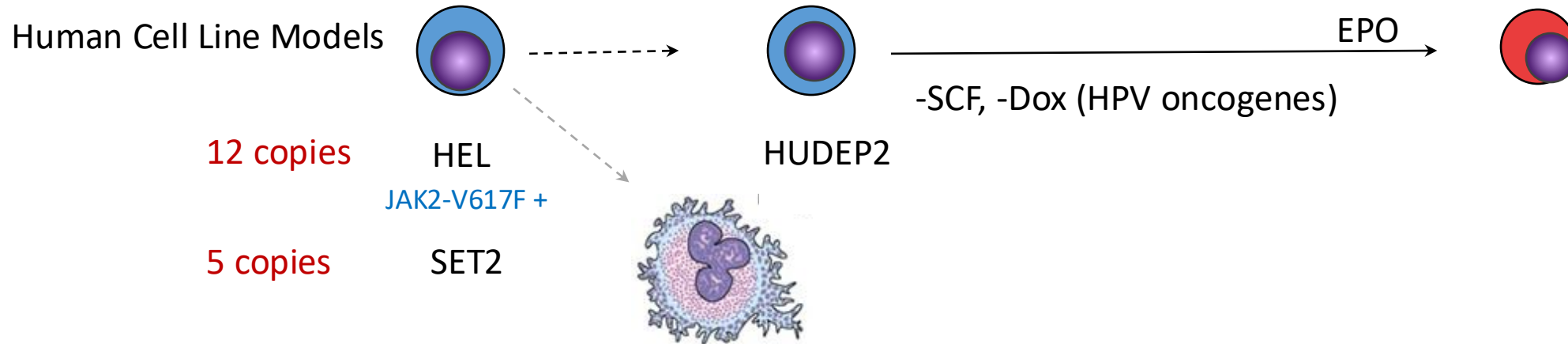
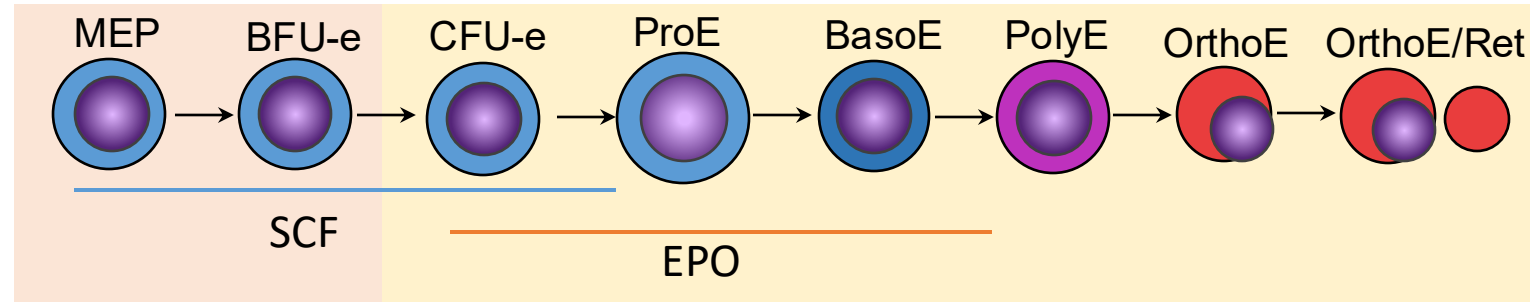
Some Research Questions worth addressing

- What is the full repertoire of STAT targets in the nucleus?
- Are there new EPOR-directed or MPL-directed biological processes to be found?
- Are the STAT targets in humans the same as in mice?
- Are there any STAT-independent transcriptional programs? How can we find them?
- Does the JAK2-V617F driver mutation induce any different transcriptional outcomes to normal signalling?

Are we just re-inventing the wheel?



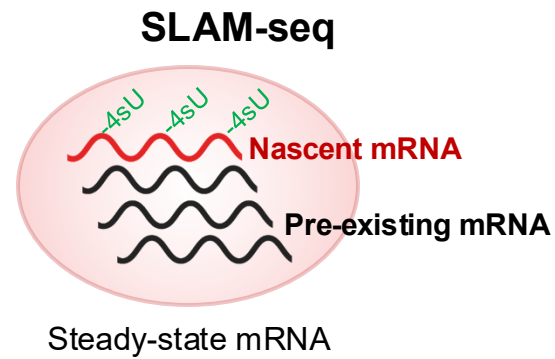
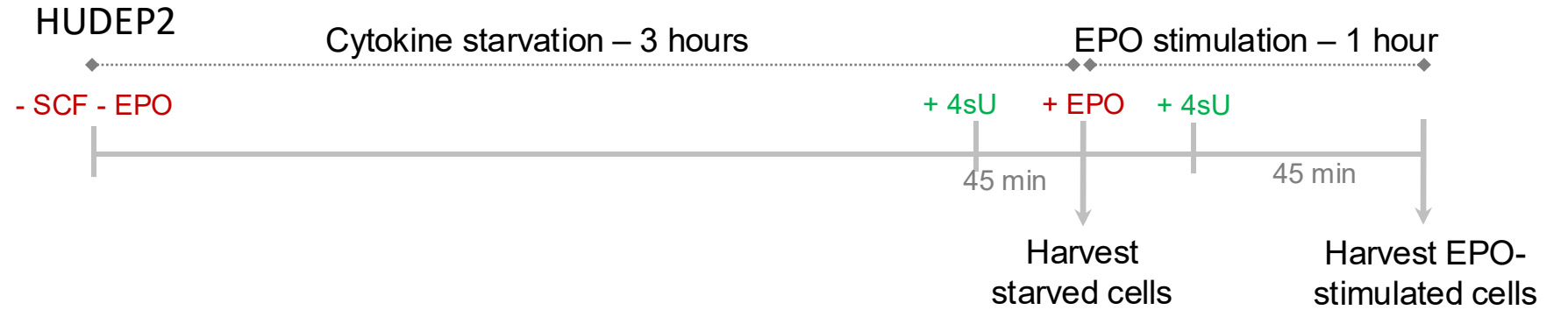
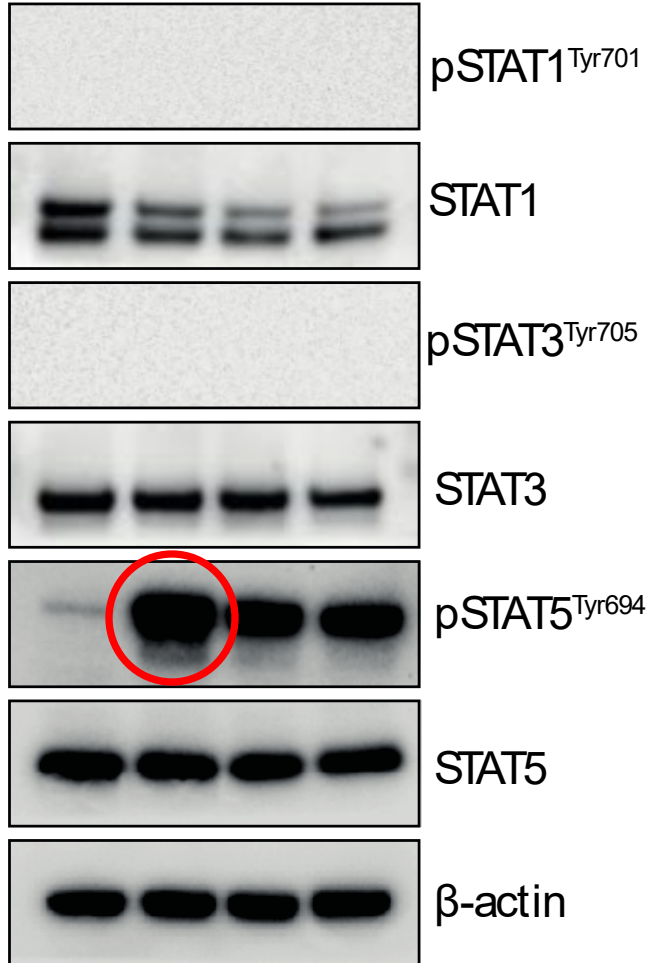
Discovery: New targets of EPOR signalling in Erythropoiesis



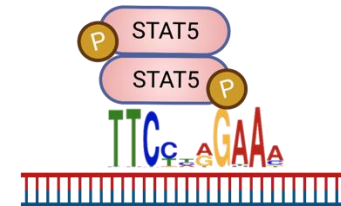
Experimental Design

EPO stimulation (hour)

0 1 2 4



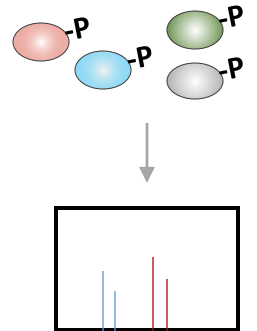
pSTAT5 ChIP-seq



ATAC-seq

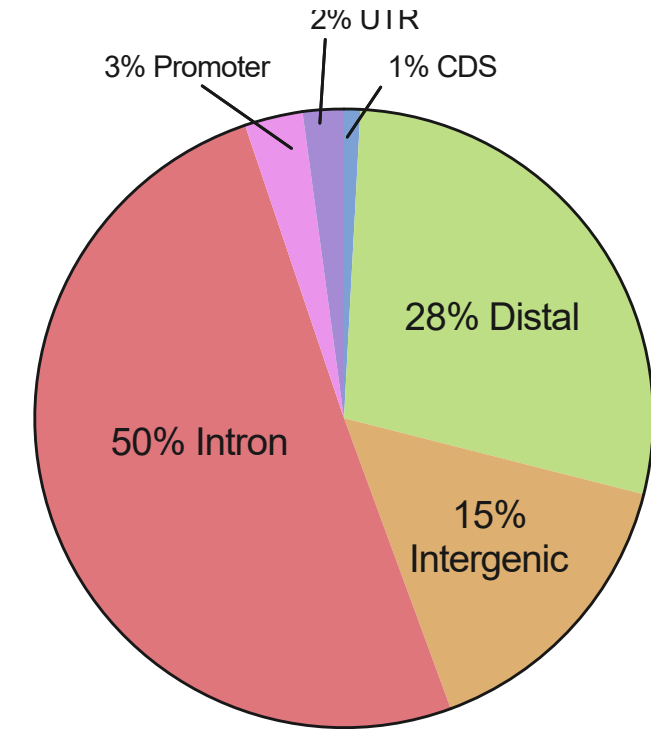
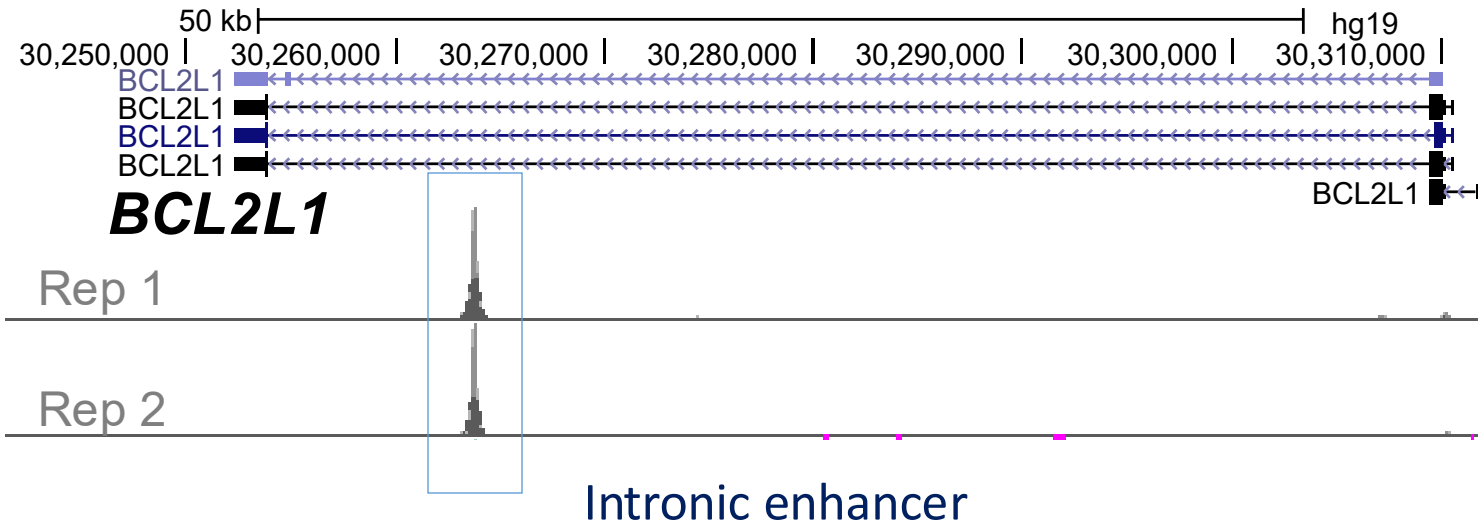
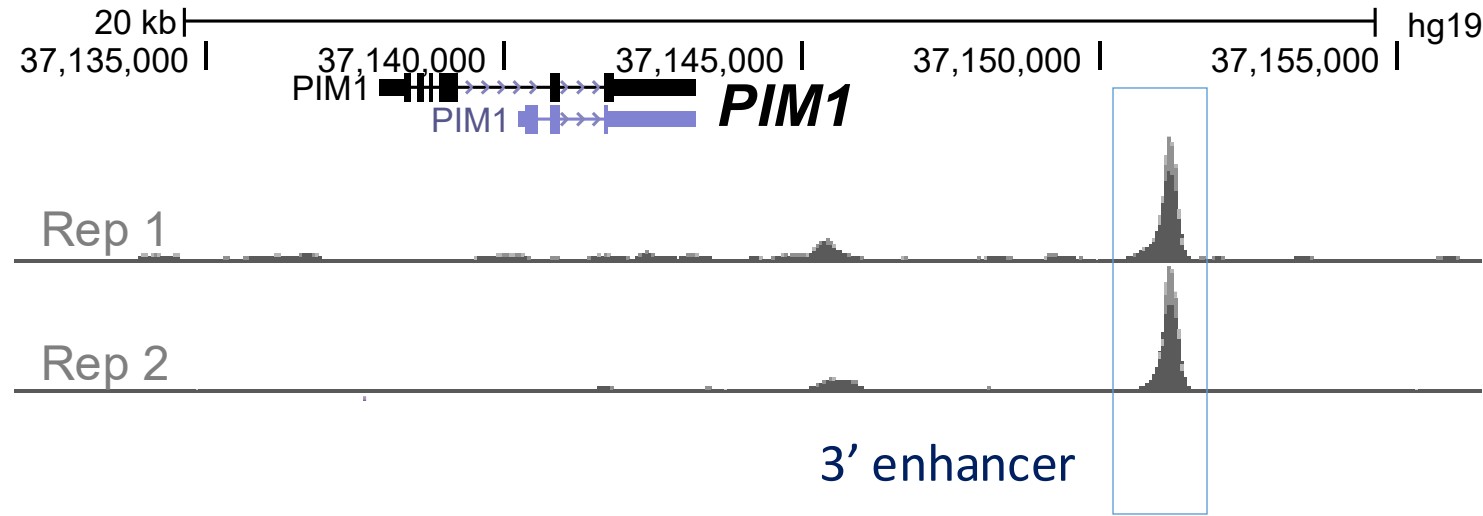


Phosphoproteomics



EPO – 5U/ml; 4sU – 400 uM

pSTAT5 binds enhancers regions at the canonical STAT-binding motif



Total = 3128



MEME
Multiple Em for Motif Elicitation

E-value = $1.2e^{-853}$



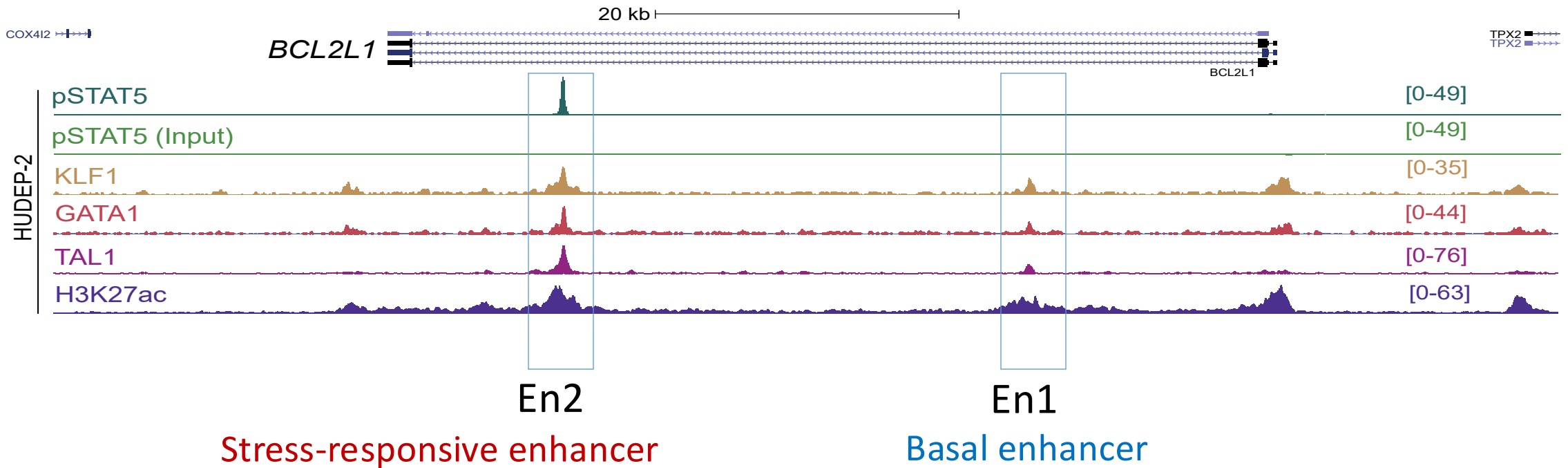
HOMER

P-value = $1e^{-1347}$



Motif present in >80% of peaks

Integration of signals from EPOR and master transcription factors at important enhancers for MPNs

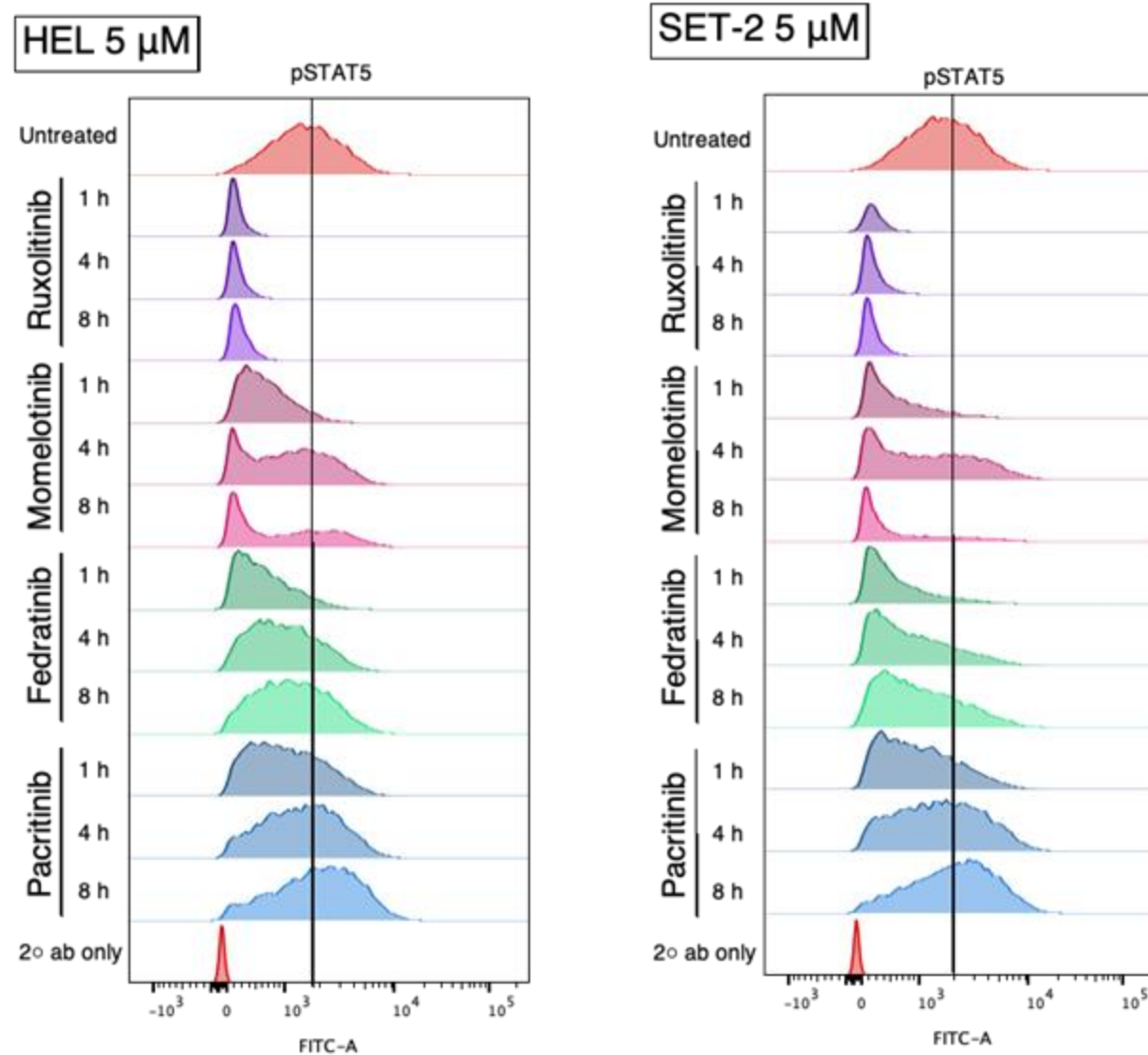


JAK inhibitors have a variety of targets

	Ruxolitinib ¹	Fedratinib ²	Pacritinib ^{2,3}	Momelotinib ²
JAK1	✓			✓
JAK2	✓	✓	✓	✓
ACVR1			✓	✓
FLT3			✓	
IRAK1			✓	
CSF1R			✓	

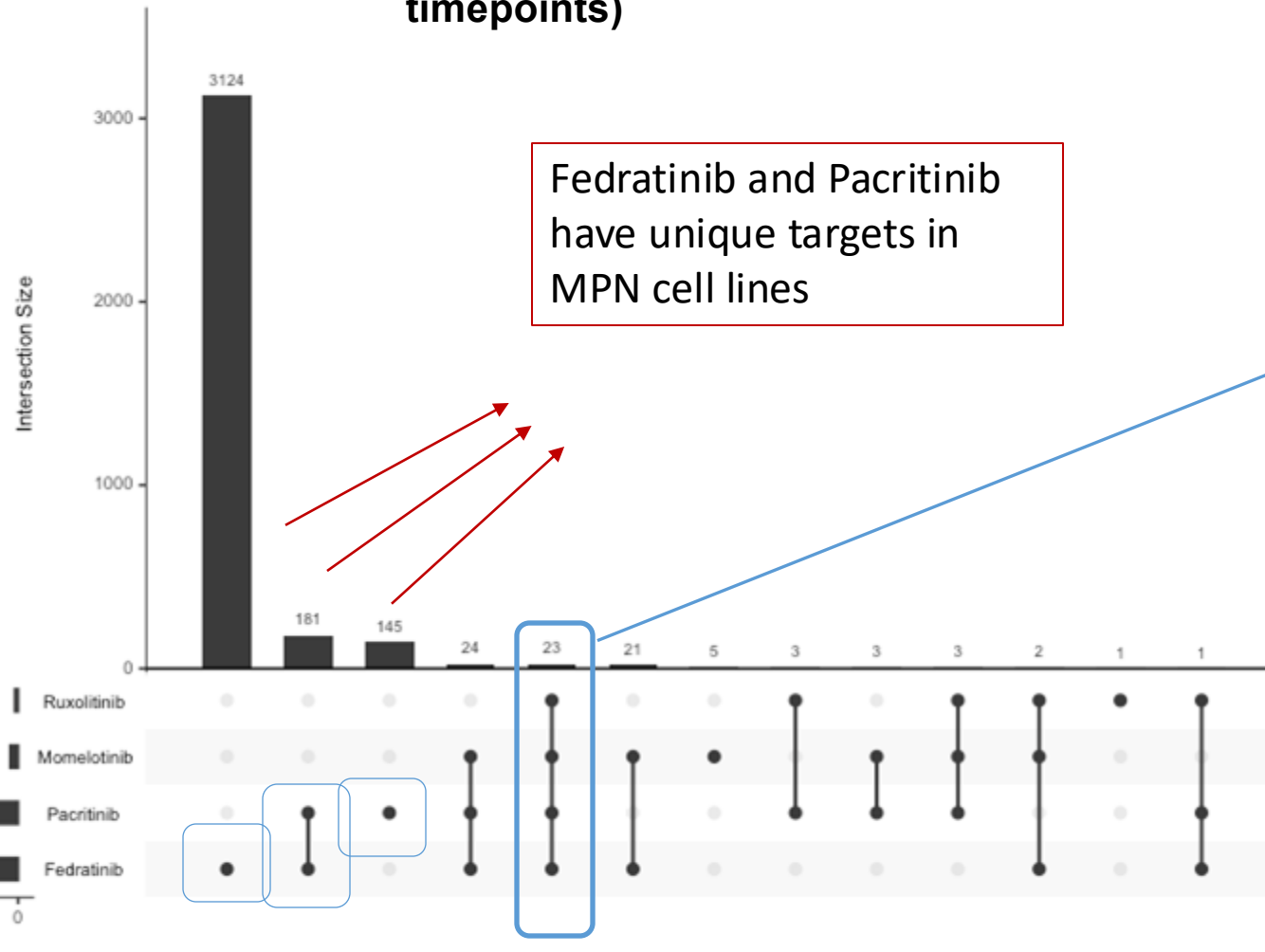
Kinase	IC50 (nM)			
	Fedratinib	Ruxolitinib	Pacritinib	Momelotinib
JAK1	105	3.3	1280	11
JAK2	3	2.8	23	18
JAK3	1002	428	520	155
TYK2	405	19	50	17

Temporal dynamics of pSTAT5 inhibition by JAK inhibitors



DRG analysis identified 23 common target genes of JAK2 inhibitors

Intersections of **DTCEs** of each JAK2 inhibitor in HEL cells (combined timepoints)

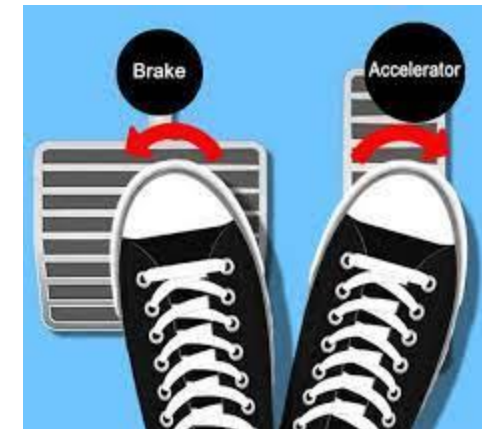


Fedratinib and Pacritinib have unique targets in MPN cell lines

EGR1
PIM2
BCL3
APOL4
NLRP3
GP6
LIMA1
GPR3
FOSL2
FOSB
ETV5
WNT11
SLC27A6
FOSL1
ID1
SOCS3
PPP1R16B
VDR
IL21R
MAFF
UBASH3B
PLCH1
AP3M2
ETV4
PPAN-P2RY11
CNN2

Pathways:

- Inflammation
- Proliferation
- Survival
- Negative regulation of JAK-STAT signalling



Lam and Perkins, unpublished data

Conclusions

- pSTAT5 binds to ~3,000 putative enhancers in human erythroid cells (some likely signalling centres for GATA1 and KLF1)
- EPO signalling induces new DNA binding activities via non-STAT TFs (e.g. EGR1)
- There are clues to new biological processes induced by EPOR signalling
- There is a quantitatively increased transcriptional response in JAK2-V617F mutant cells but no new genes were identified

Can we improve survival in MF?

- Allogeneic BMT remains the only curative therapy in MF
- Ruxolitinib improves survival, but does not cure MF
- Potential **disease modifying agents** in trials (need well controlled up front trials with long end points (OS) or surrogate end points for OS (allele burden))

Interferon- α

BET inhibitors

Telomerase inhibitors

Combination therapies: PI3K inhibitors, PIM inhibitors, ERK inhibitors

Back to the drawing board....finding new targets

- Have a look at the draft recommendations for New Guidelines on Myelofibrosis

Myelofibrosis

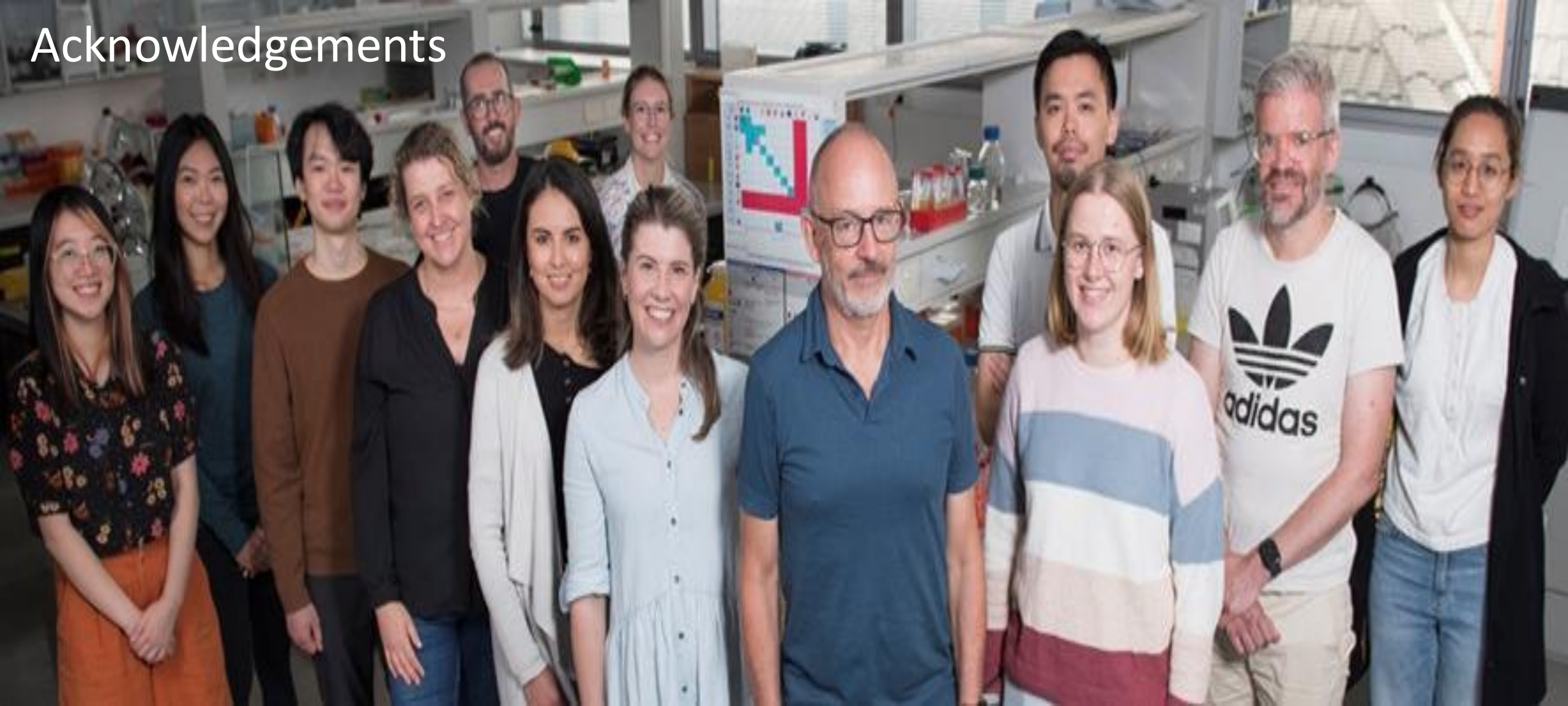
ASH Seeks Public Comment on Draft Recommendations for New Guidelines on Myelofibrosis

ASH is now accepting public comments on draft recommendations on myelofibrosis. Comments will be accepted until November 24, 2025. We encourage all interested healthcare professionals, stakeholder organizations, and individuals to share their feedback on these draft recommendations. All comments will be reviewed by the guideline panel before the guidelines are finalized.

MYELOFIBROSIS DRAFT RECOMMENDATIONS

Please submit all comments to guidelines@hematology.org.

Acknowledgements



Contacts

Clinical Genomics

Trials

Research Genomics

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